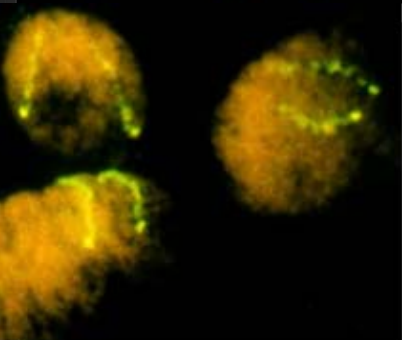
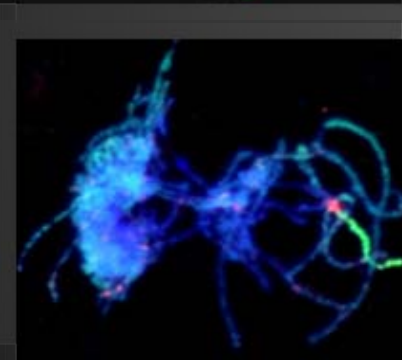
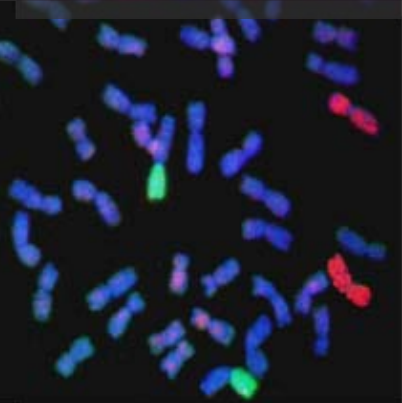
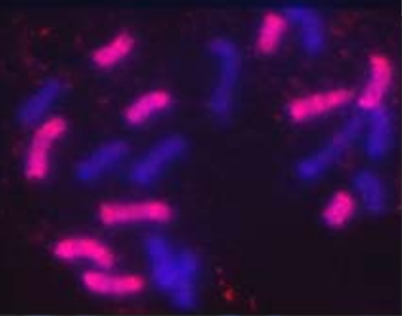


# Genomic *in situ* hybridization to understand hybrid genomes

Trude Schwarzacher  
Department of Biology  
University of Leicester, UK

[ts32@le.ac.uk](mailto:ts32@le.ac.uk)

[www.molcyt.com](http://www.molcyt.com)



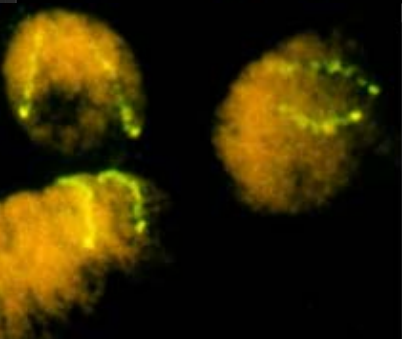
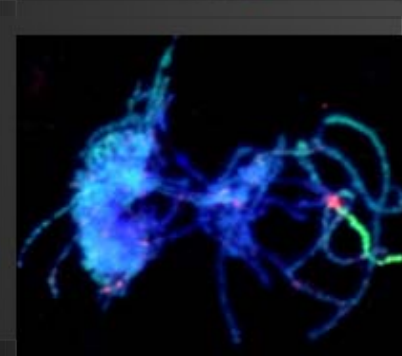
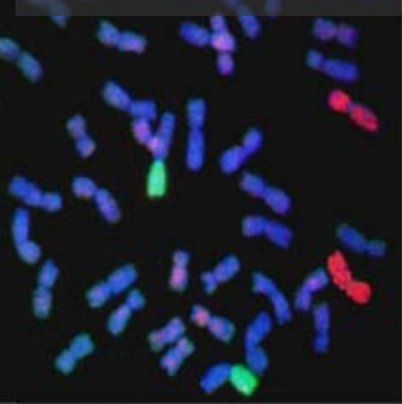
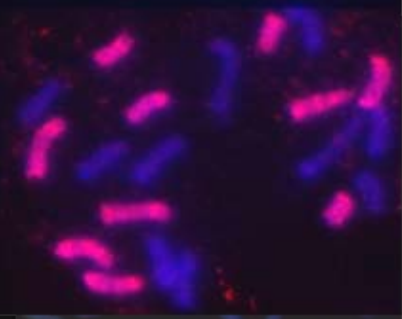
# GENOMIC *IN SITU* HYBRIDIZATION GISH

## ◎ Background and history

- What is it?
- What can it do?

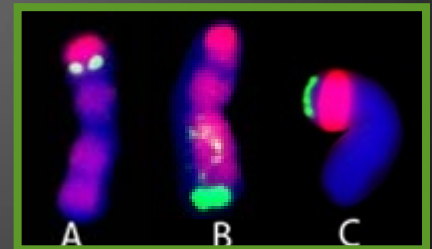
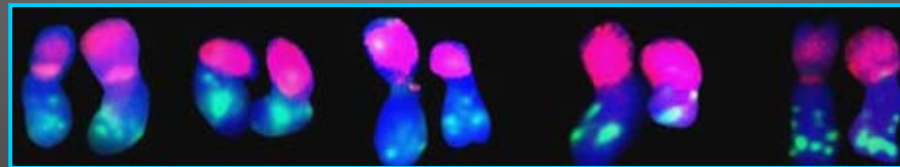
## ◎ Applications

- Examples from our lab
  - Wheat
  - Brassica
  - Crocus



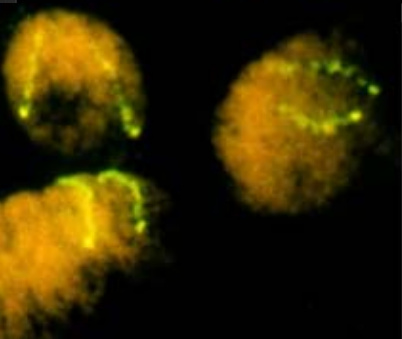
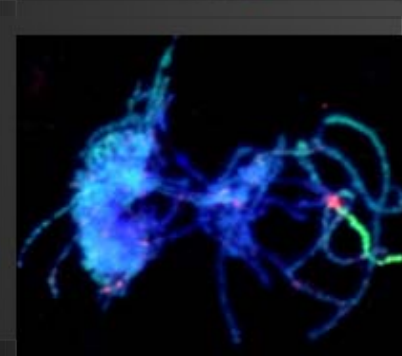
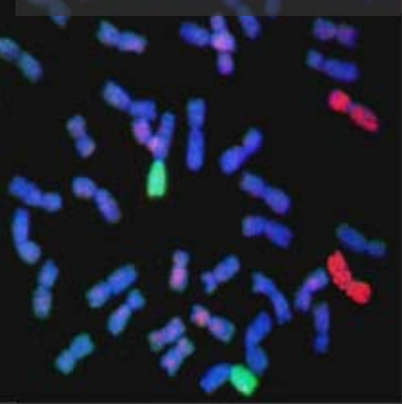
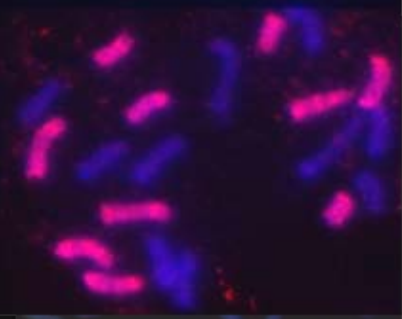
# GENOMIC *IN SITU* HYBRIDIZATION GISH

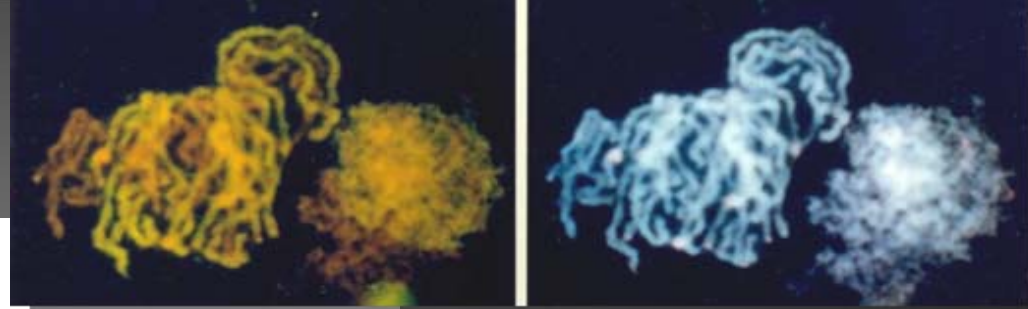
- Uses total genomic DNA as a probe for *in situ hybridization* to chromosomes and nuclei
- Identifies origin of chromatin
- Parental origin of hybrids
  - Can be combined with other probes, most often repetitive DNA probes
- Alien chromatin in breeding lines
  - Size and origin



# GENOMIC *IN SITU* HYBRIDIZATION

- Parental origin of hybrids
  - Auto or allo-poliploidy
- Alien chromatin in breeding lines
  - Size and origin
  - Recipient chromosome
- Meiosis and chromosome pairing
- Interphase cytogenetics



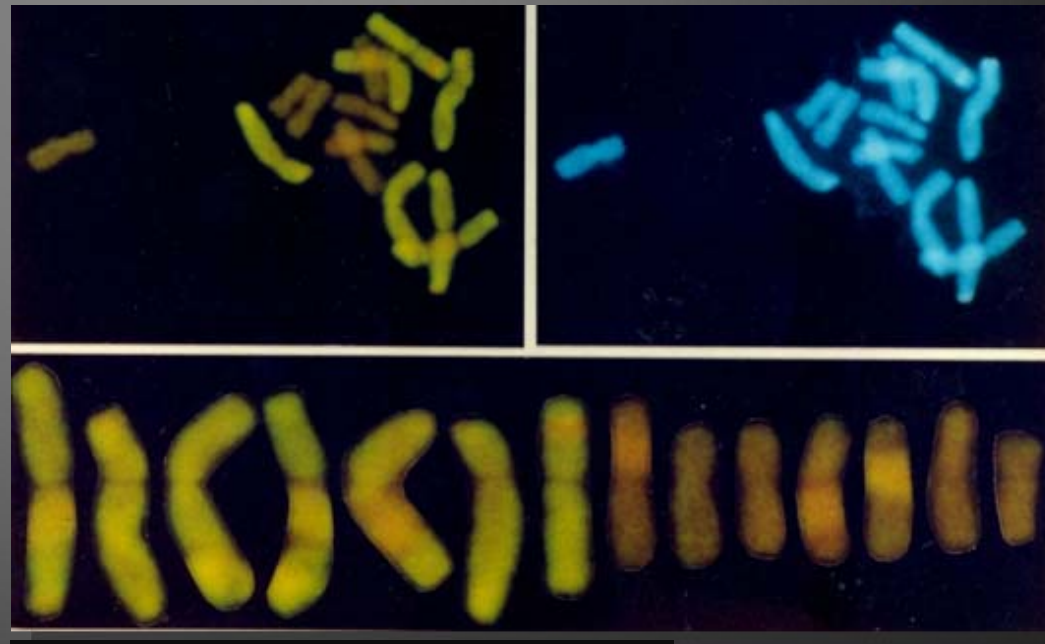


## ***In Situ* Localization of Parental Genomes in a Wide Hybrid**

T. SCHWARZACHER\*, A. R. LEITCH, M. D. BENNETT†  
and J. S. HESLOP-HARRISON

*Cambridge Laboratory, Institute of Plant Science Research, Trumpington, Cambridge CB2 2JB, UK*

Accepted: 7 March 1989

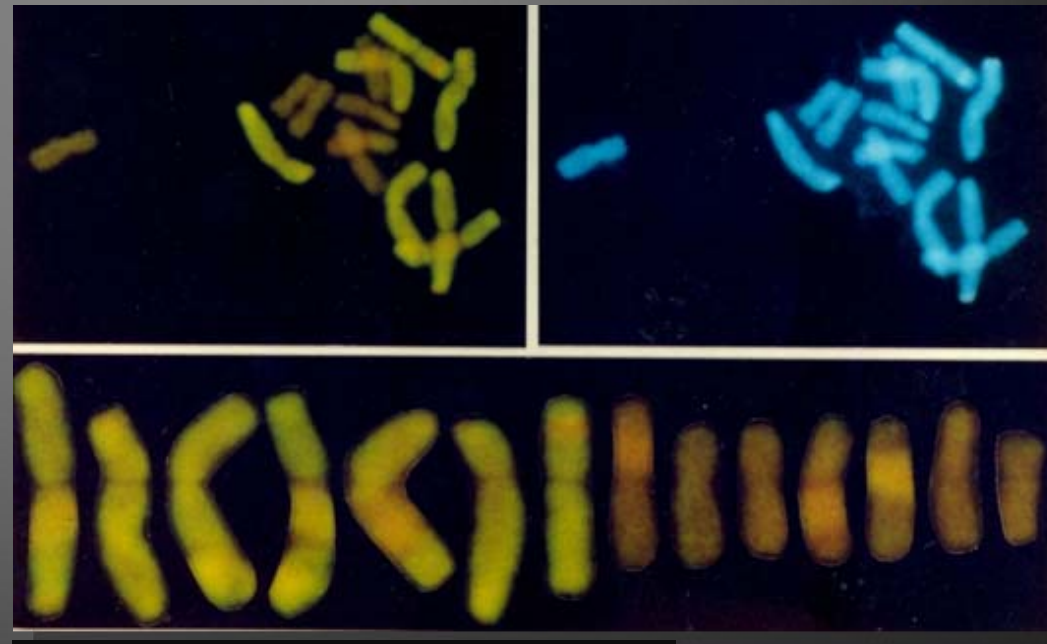
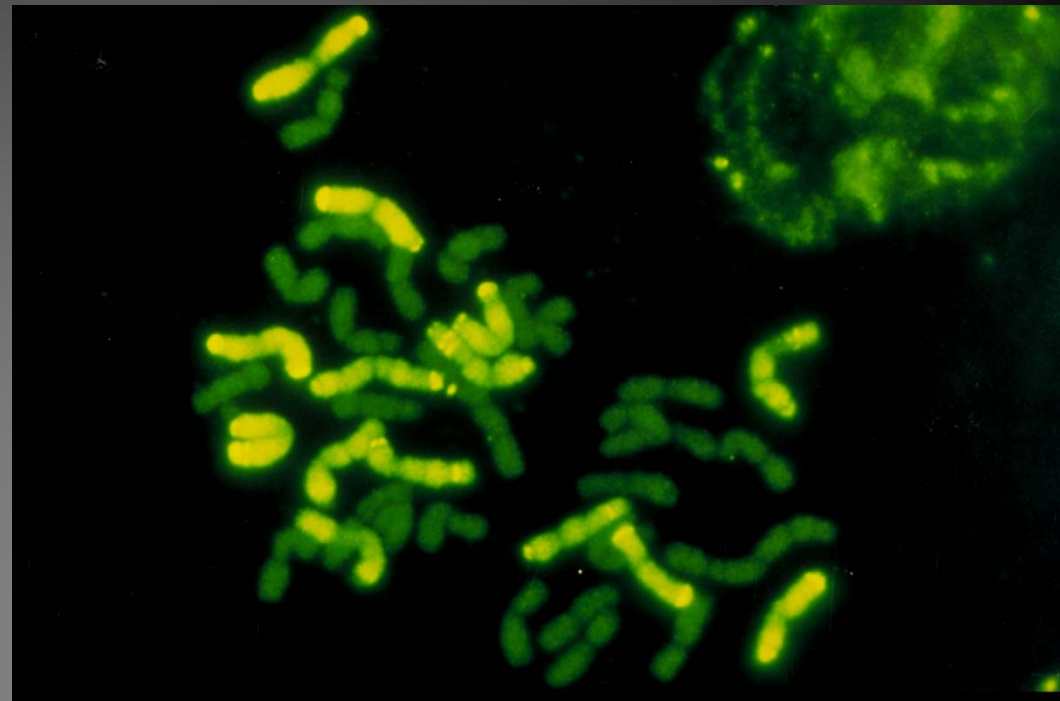




Addition of unlabelled,  
cold total genomic DNA  
to block common  
sequences between the  
parental genomes

Heslop-Harrison, J.S., Schwarzacher,  
T., Leitch, A.R., Anamthawat-  
Jonsson, K. and Bennett, M.D. (1988)  
A method of identifying DNA sequences in  
chromosomes of plants. European Patent  
Application 8828130.8. December 8, 1988.  
Early Publication, June 8, 1990.

Anamthawat-Jonsson, K.,  
Schwarzacher, T., Leitch, A.R.,  
Bennett, M.D. and Heslop-Harrison,  
J.S. (1990)  
Discrimination between closely related  
Triticeae species using genomic DNA as a  
probe. Theoretical and Applied Genetics 79,  
721-728.



## Genomic in situ hybridization to identify alien chromosomes and chromosome segments in wheat

**T. Schwarzacher<sup>1</sup>, K. Anamthawat-Jónsson<sup>1,2</sup>, G. E. Harrison<sup>1</sup>, A. K. M. R. Islam<sup>3</sup>, J. Z. Jia<sup>4</sup>, I. P. King<sup>1</sup>, A. R. Leitch<sup>1,5</sup>, T. E. Miller<sup>1</sup>, S. M. Reader<sup>1</sup>, W. J. Rogers<sup>1,6</sup>, M. Shi<sup>1</sup> and J. S. Heslop-Harrison<sup>1</sup>**

<sup>1</sup> John Innes Centre for Plant Science Research, Colney Lane, Norwich NR4 7UJ, UK

<sup>2</sup> Agricultural Research Institute, Keldnaholt, Reykjavik, 112, Iceland

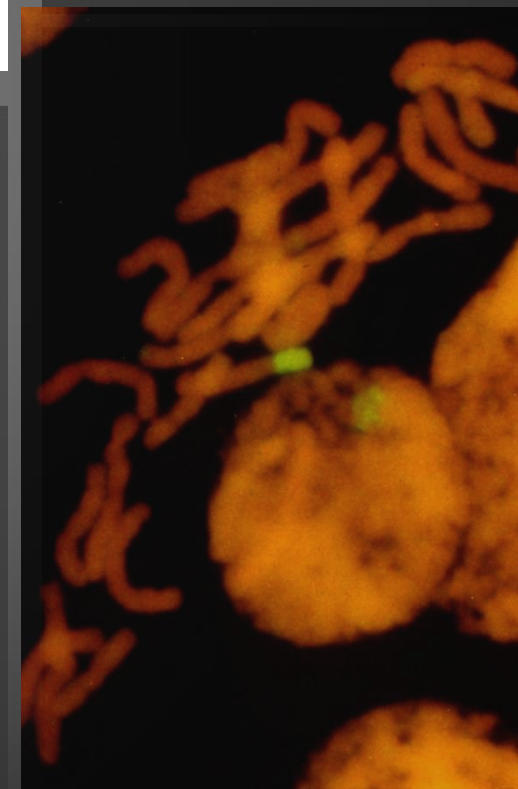
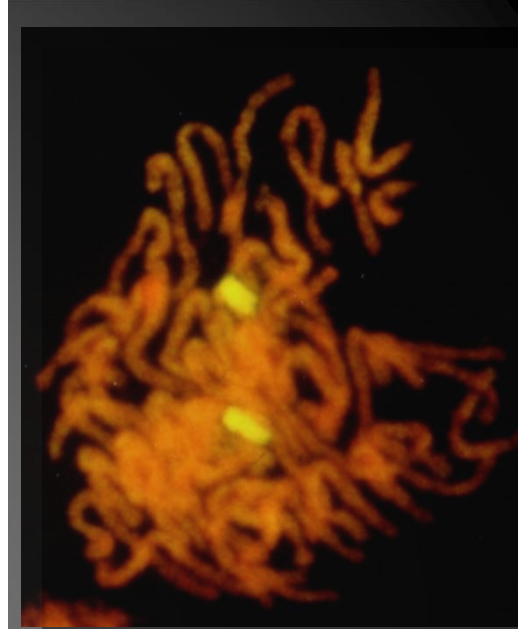
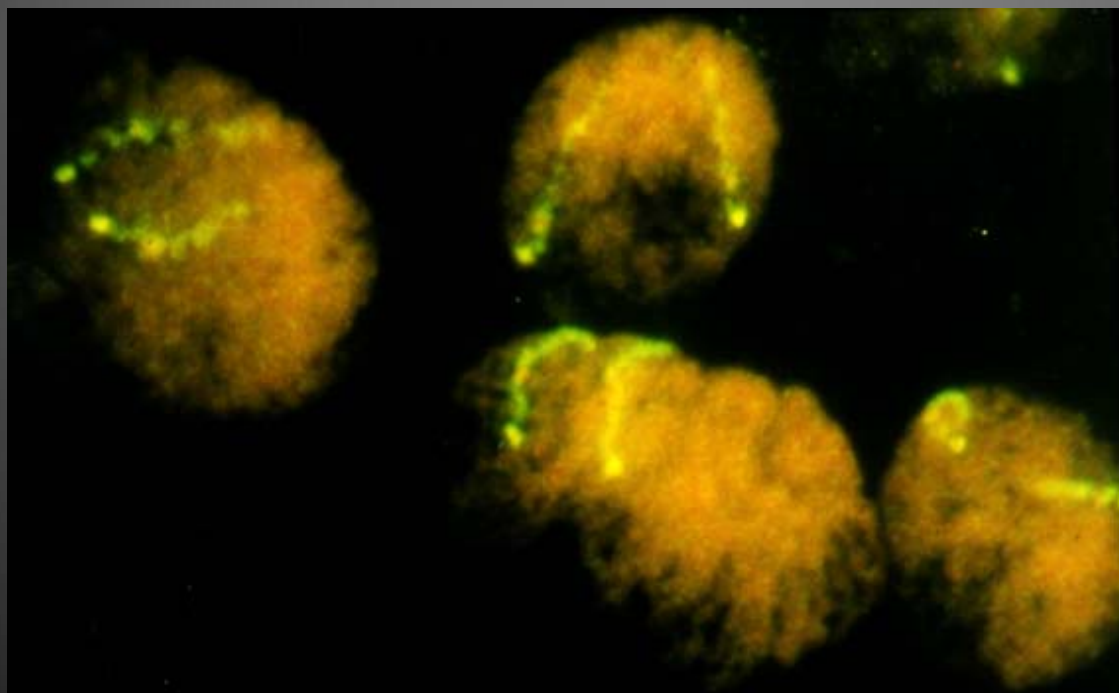
<sup>3</sup> Waite Institute, University of Adelaide, Glen Osmond SA 5064, Australia

<sup>4</sup> The Institute of Crop Germplasm Resources, Chinese Academy of Agricultural Sciences, Beijing 100081, China

<sup>5</sup> Queen Mary and Westfield College, Mile End Road, London E1 4NS, UK

<sup>6</sup> School of Life Sciences, University of Liverpool, P.O. Box 147, Liverpool L69 3BX, UK

Received December 12, 1991; Accepted February 26, 1992



# EXAMPLES WHERE GISH HAS BEEN SUCCESSFUL

- Triticae
  - *Triticum, Aegilops, Hordeum, Haynaldia, Thinopyrum, Secale, Hystrix, Leymus, Agropyron, Elymus, Elytrigia*
- *Brachypodium*
- *Oryza* genomes
- *Zea mays*
- *Pennisetum*
- *Tripsacum*
- *Saccharum*
- *Avena*
- *Lolium* and *Festuca* hybrids *Festulpia*
  
- *Eleusine*
- *Alstroemeria*
- *Aloe*
- *Lilium*
- *Allium*
- *Crocus*
- *Tulipa*
- *Musa* genomes
  
- Asteraceae
  - *Dahlia*
  - *Chrysanthemum, Dendranthema and Argyranthemum*
- Brassicaceae
  - Brassica species, alien and hybrids with *Eruca, Orchyophragmus, Sinapis, Raphanus Brassica, Lesquerella fendleri, Arabidopsis* species/hybrids
- Solanaceae
  - *Solanum*, potato, tomato
- Fabaceae
  - *Arachis*
  - *Medicago*
- *Coffea arabica*
- *Gossypium*
- *Rubus*
- *Beta*
- *Zingeria*
- *Setaria*
  
- *Phalaenopsis*



# *Callitriche platycarpa* (2n=20)

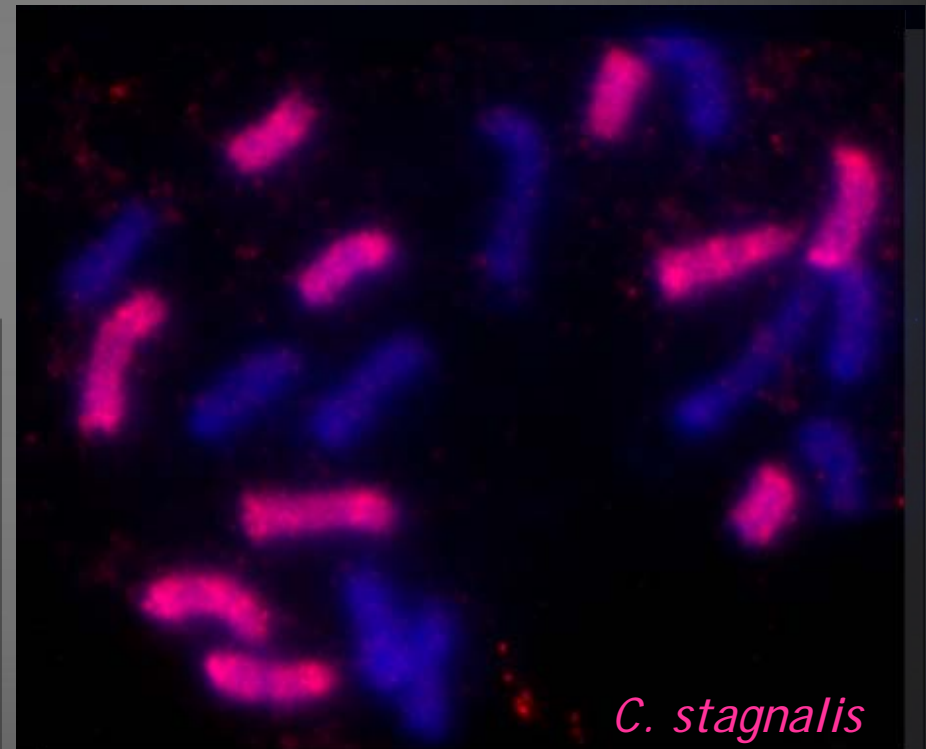
## Water starworts



Savidge (1960): *allo-tetraploid* origin from *C. stagnalis* and *C. cophocarpa* (both 2n=10)

Schotsman (1967):  
~~auto-tetraploid~~ from  
*C. cophocarpa*

British Species	2n
<i>Callitriche hermaphroditica</i> L.	6
<i>Callitriche truncata</i> Guss.	6
<i>Callitriche stagnalis</i> Scop.	10
<i>Callitriche obtusangula</i> Le Gall	10
<i>Callitriche platycarpa</i> Kütz.	20
<i>Callitriche palustris</i> L.	20
<i>Callitriche brutia</i> Petagna	28
<i>Callitriche hamulata</i> Kütz.	38



# *Crocus* species and hybrids



*C. flavus*  
 $2n=8$

B.J. 94



*C. 'Stellaris'*  
 $2n=2x=10$



*C. angustifolius*  
 $2n=12$



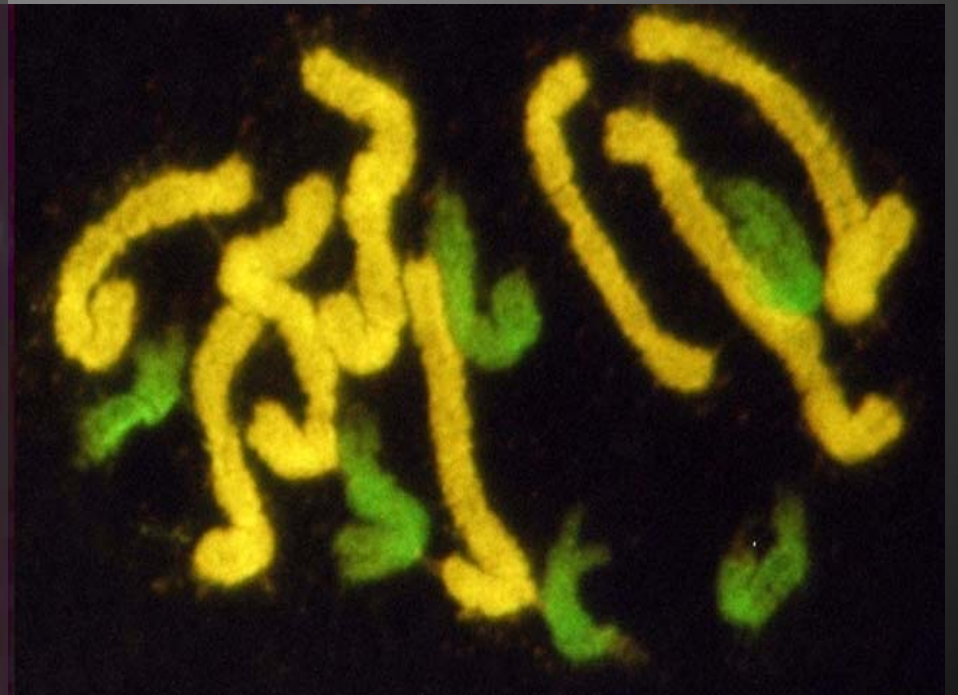
*C. 'Golden Yellow'*  
 $2n=3x=14$

# 'GOLDEN YELLOW' CROCUS

$2n=3x=14$

6 chr from *Crocus angustifolius*

2x4 chr from *Crocus flavus*





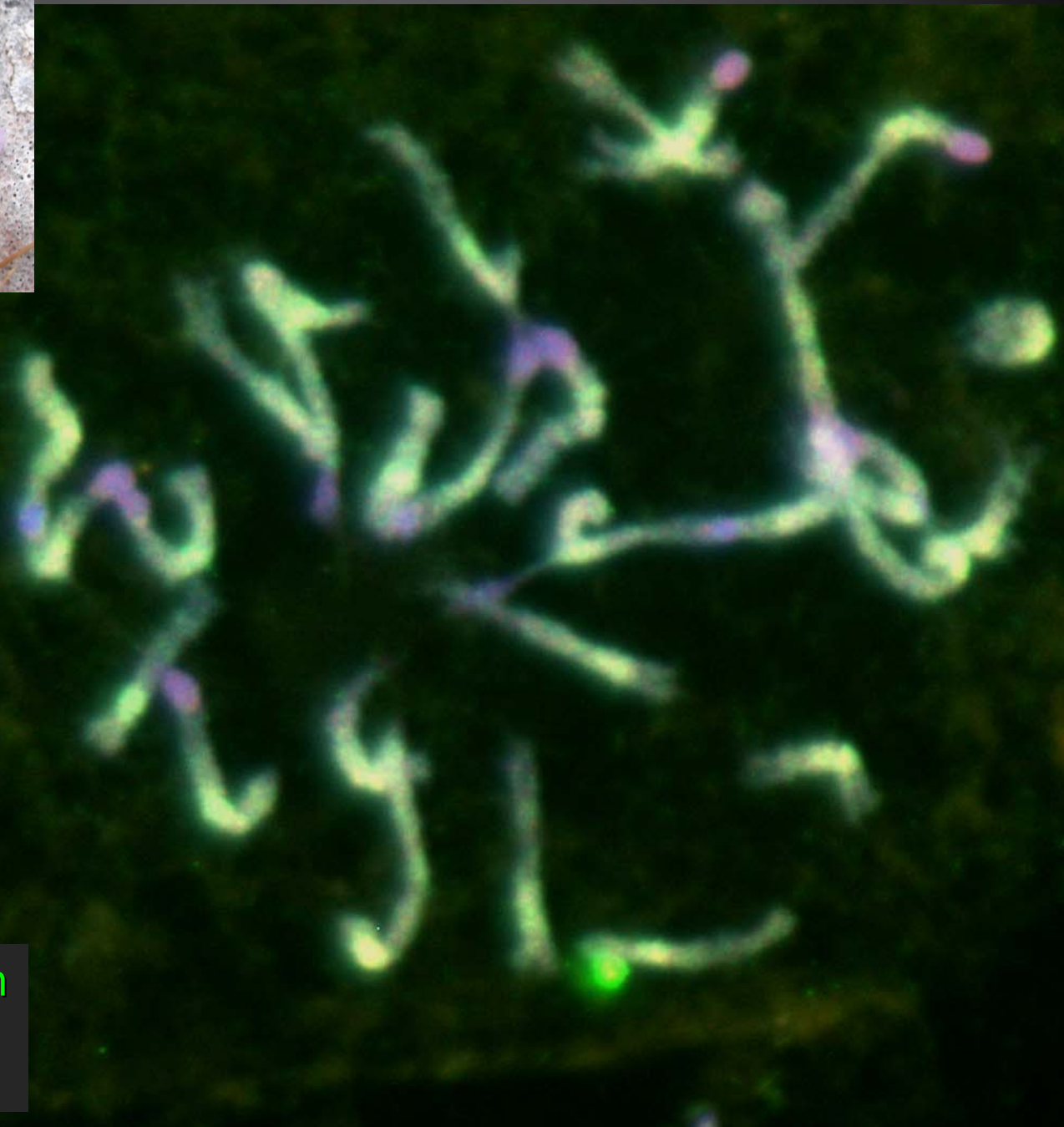


[www.CrocusBank.org](http://www.CrocusBank.org)





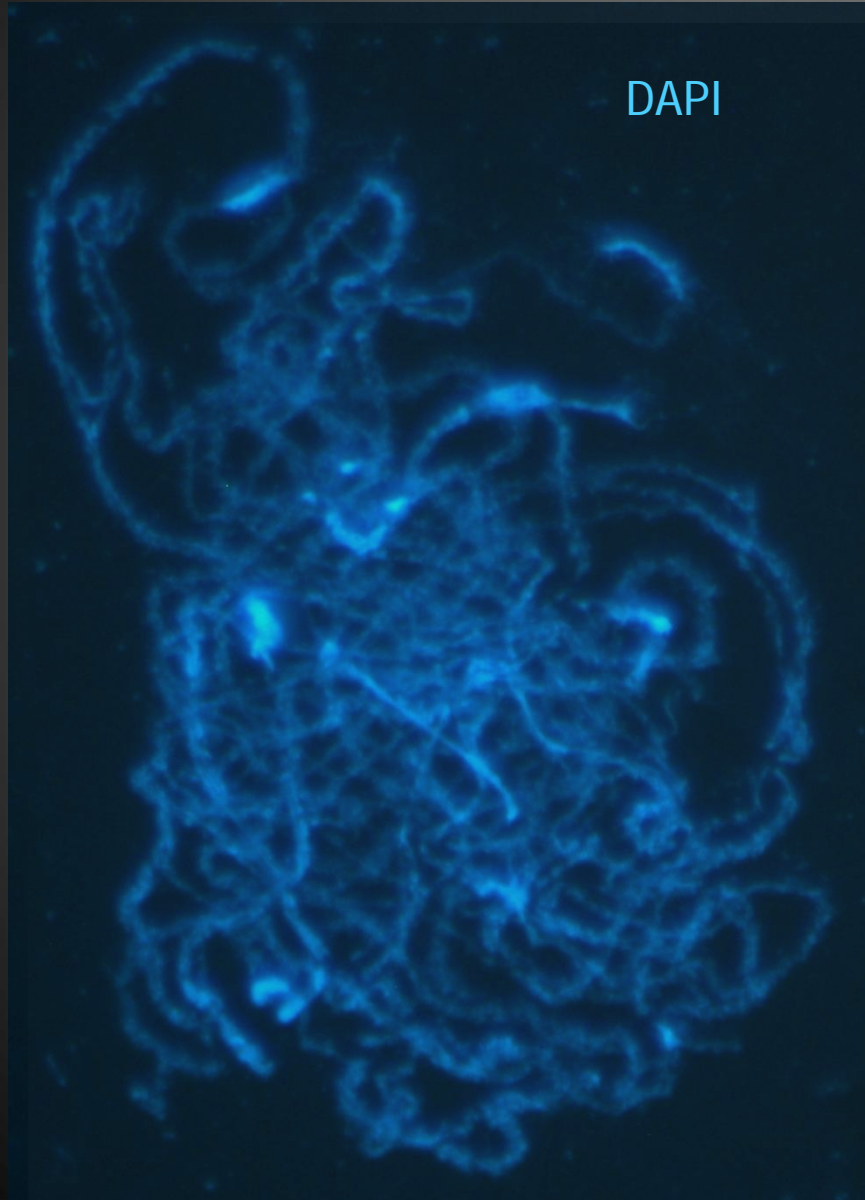
Saffron  
*Crocus sativa*  
 $2n=3x=24$



*C. cartwrightianus* green

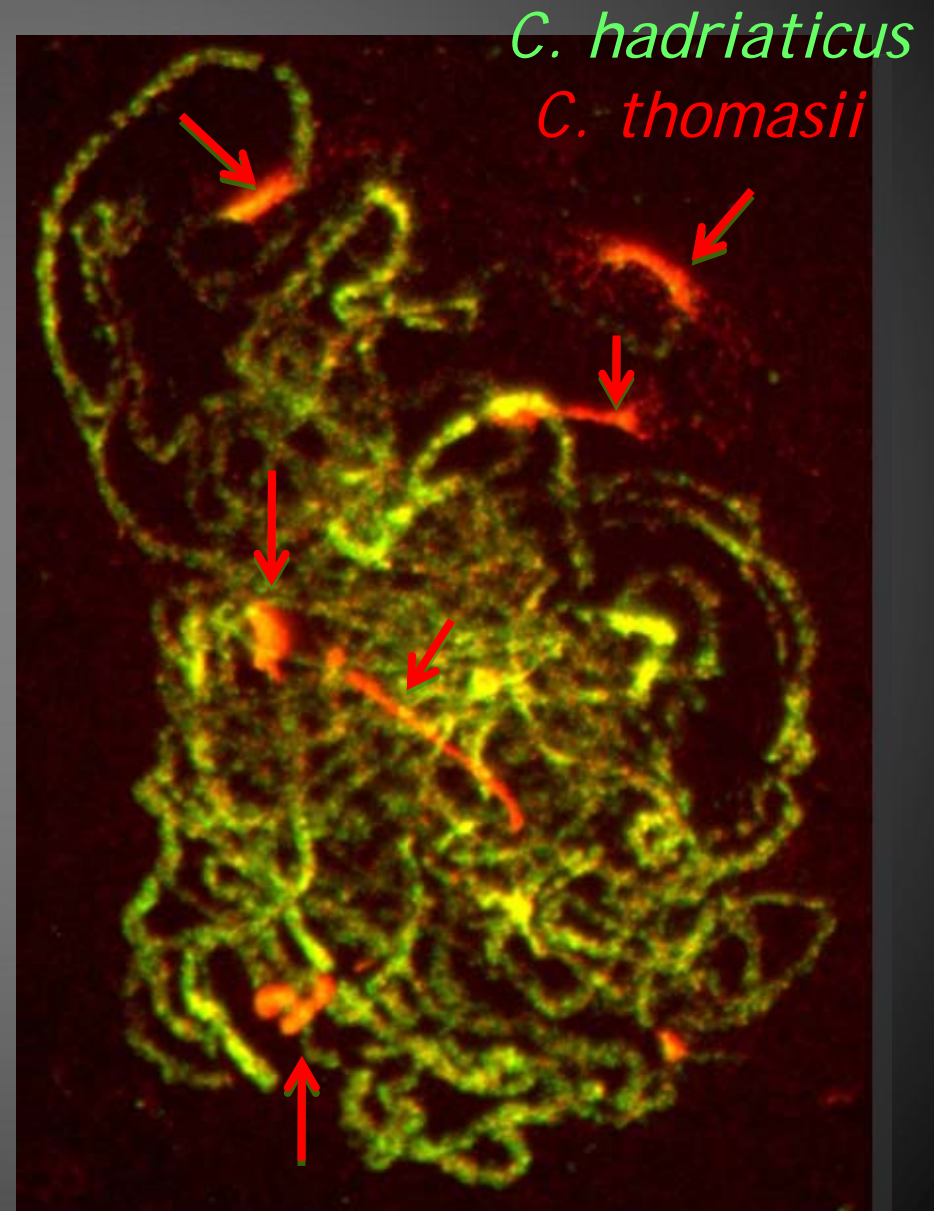
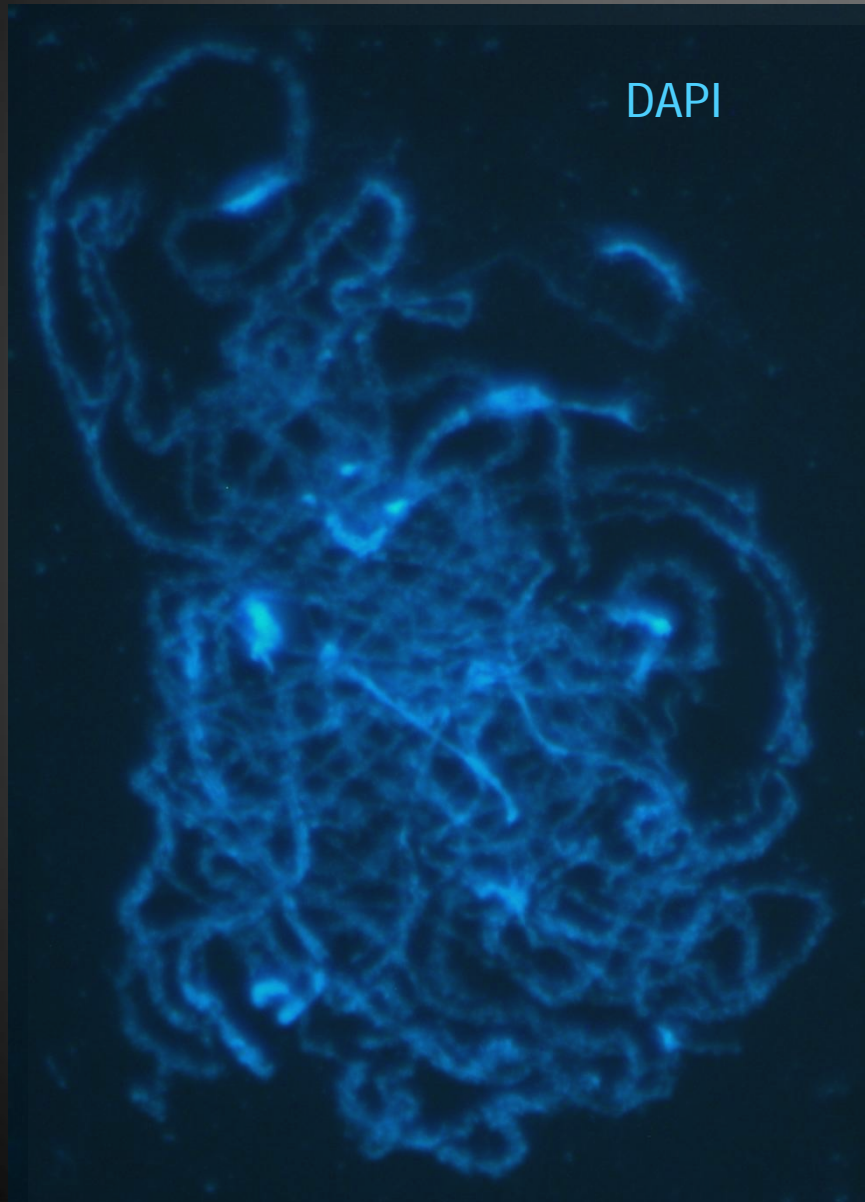
*C. thomasii* red

# *Crocus sativus* Pachytene

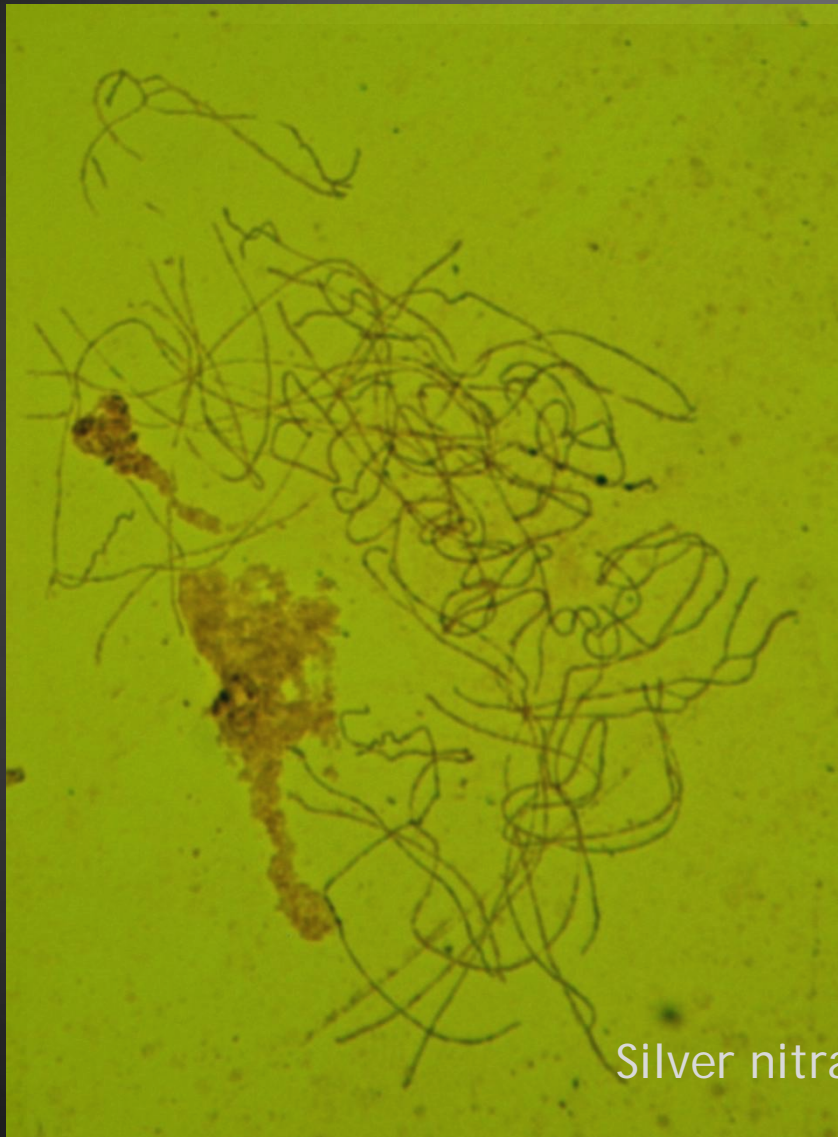




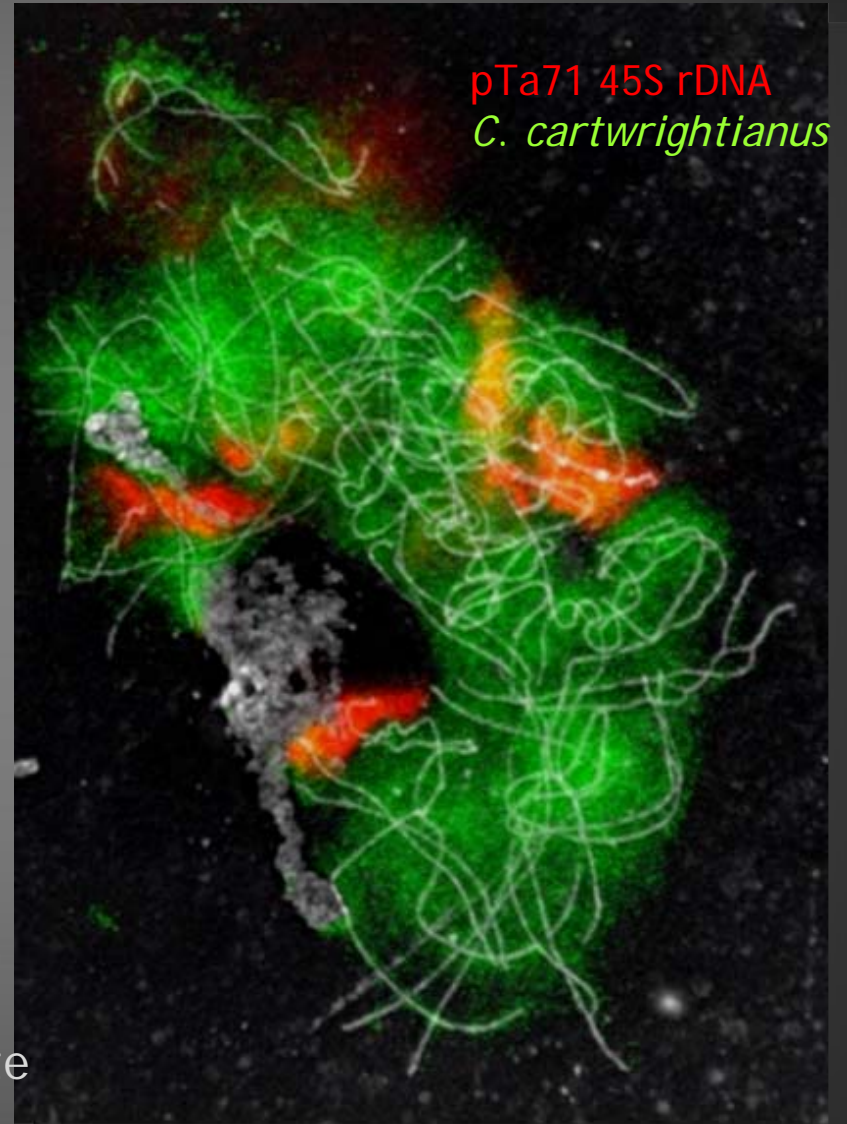
# *Crocus sativus* Pachytene



# *Crocus sativus* Pachytene



Silver nitrate



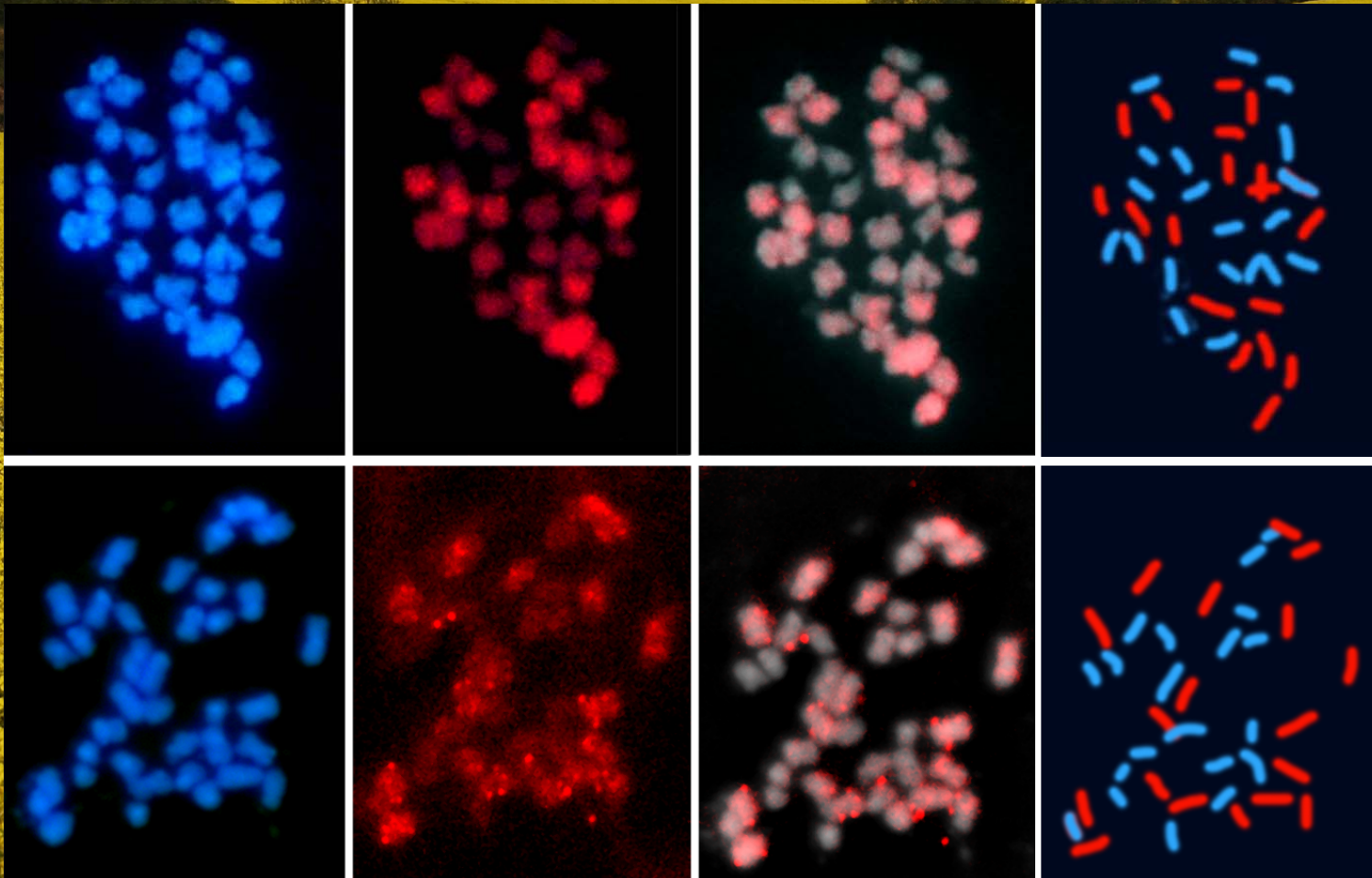
pTa71 45S rDNA  
*C. cartwrightianus*



Rapeseed *B. napus* (AACC,  $2n=4x=38$ ) - hybridized with C-genome CACTA element red

*B. oleracea* (CC,  $2n=2x=18$ )

*B. rapa* (AA,  $2n=2x=20$ )

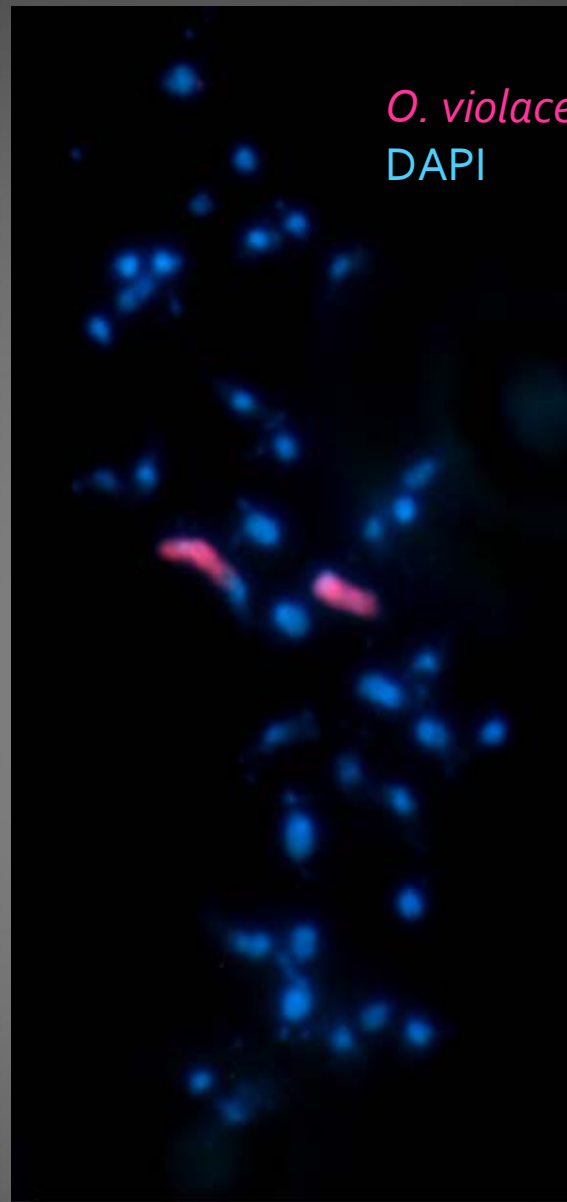


Pat Heslop-Harrison  
Karine Alix  
Xianhong Ge

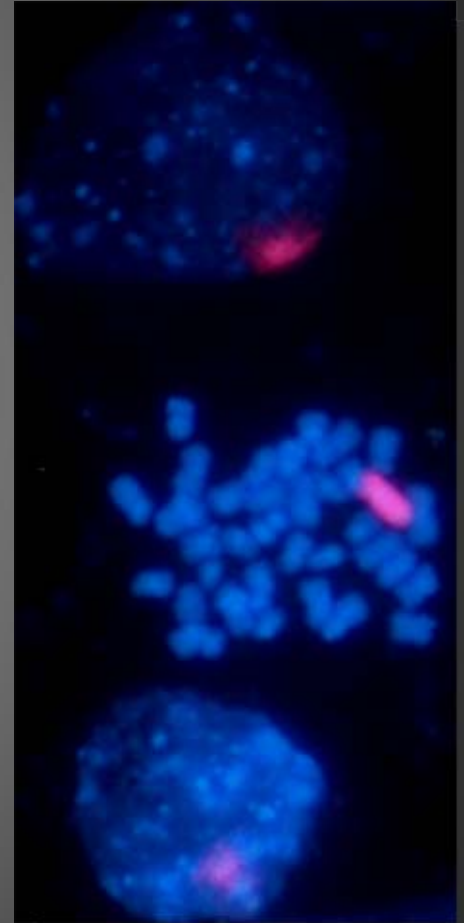
Genome Specificity of a CACTA (*En/Spm*) Transposon

*Brassica*  
synthetic  
hexaploids

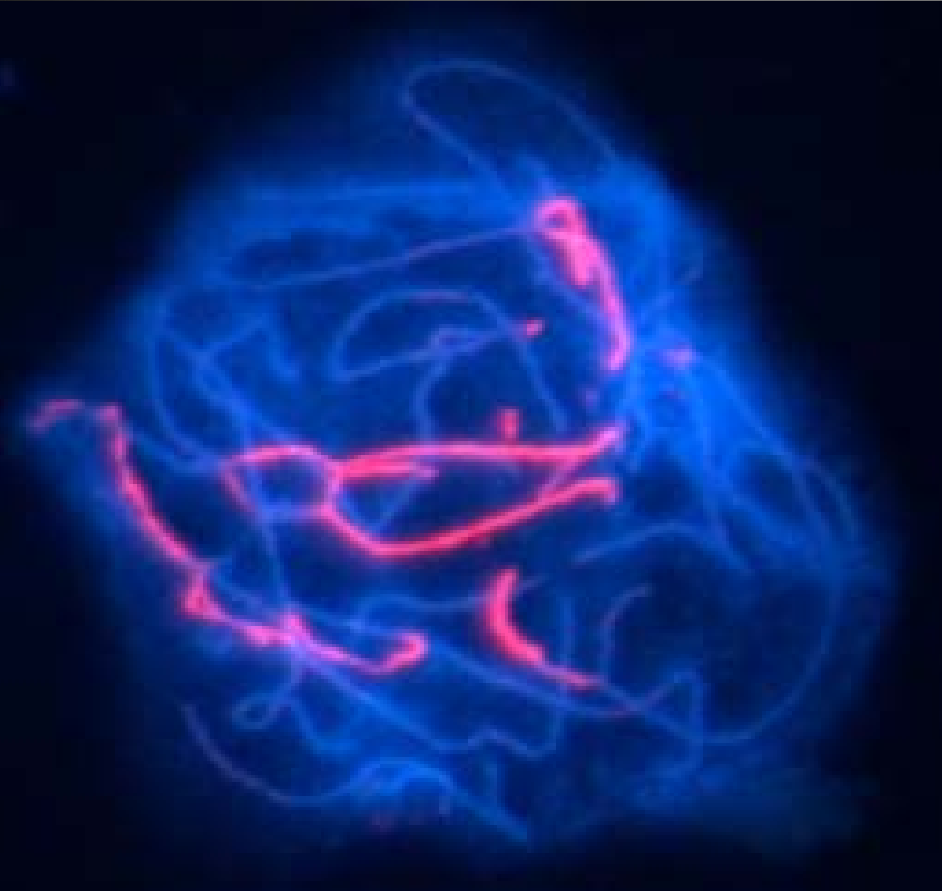
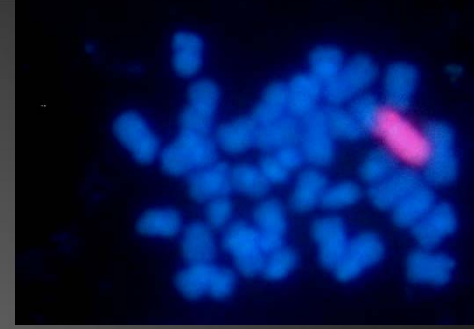
*Brassica napus*  
addition line  
*Orychophragmus*  
*violaceus*



*O. violaceus* genomic DNA  
DAPI



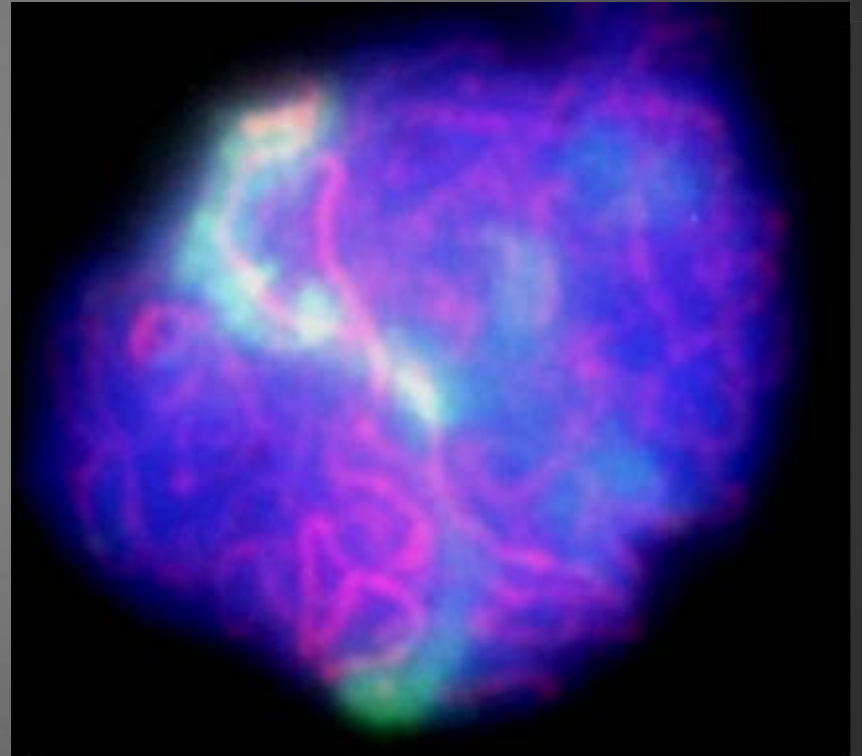
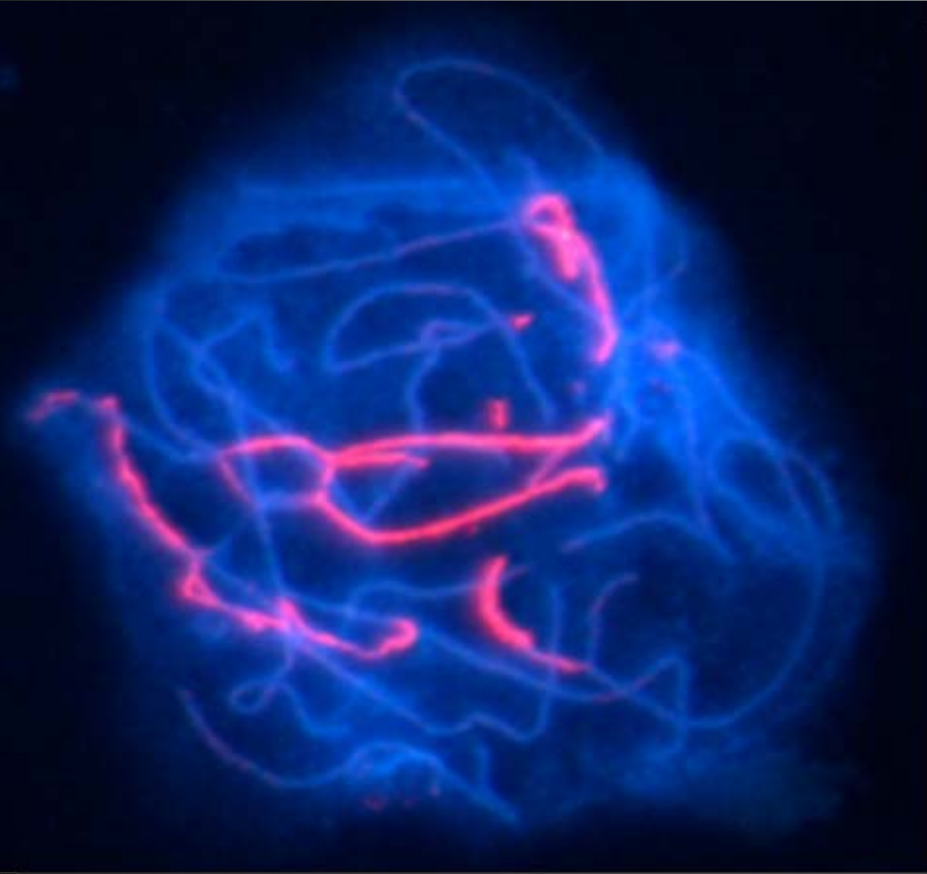
**Brassica Monosomic  
Addition Line**  
*Orychophragmus violaceus*



DAPI, ASY-1



# Brassica Monosomic Addition Line *Orychophragmus violaceus*



DAPI, ASY-1 *O. violaceus* genomic DNA  
partly unpaired and partly paired with Brassica chromosomes



# Alien gene transfer, alien recombinant chromosomes



# Total genomic DNA can be used as a probe to distinguish



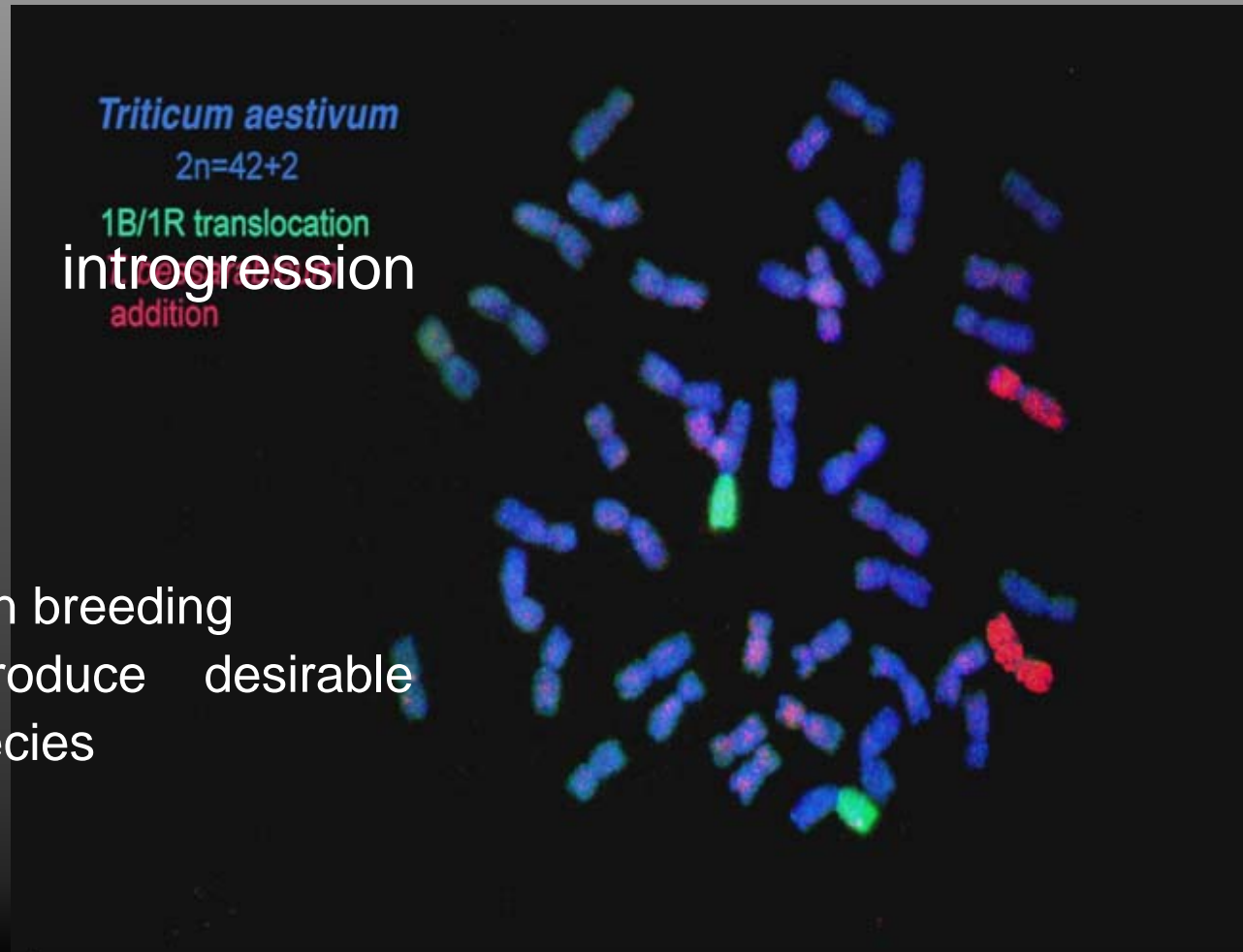
- Genomes in sexual hybrids

- Alien chromosome

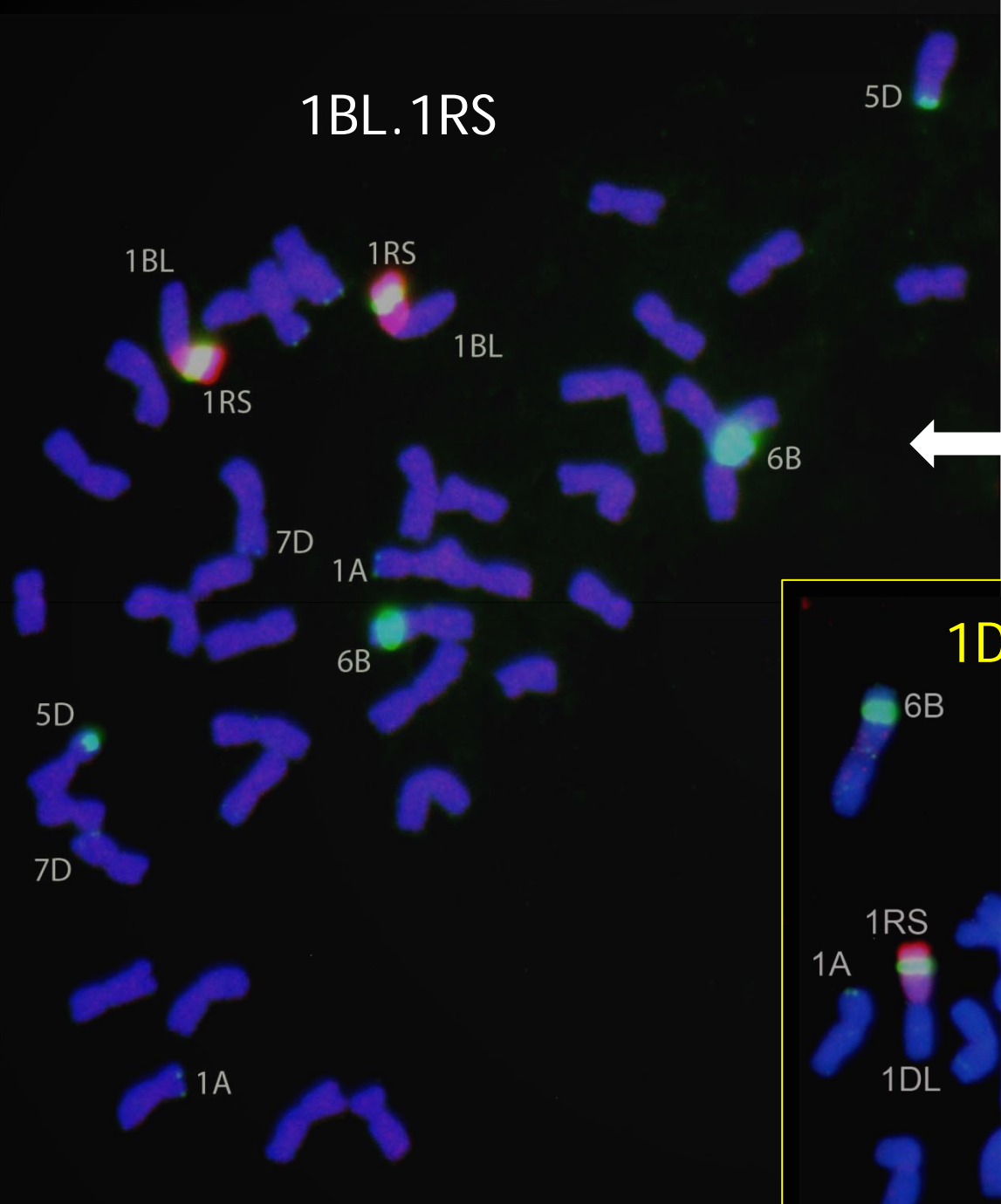
- Additions

- Translocations

- Used extensively in breeding programmes to introduce desirable traits from wild species



1BL.1RS



Rye DNA

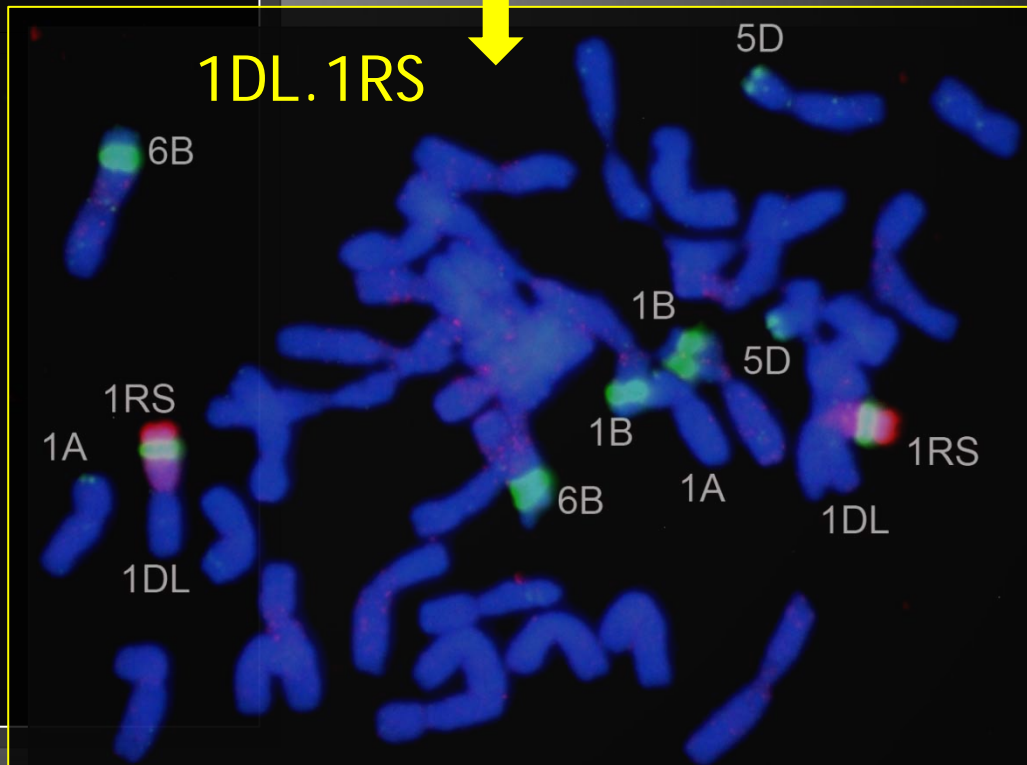
pTa71-45S rDNA

4 major sites 1RS, 6BS

6 major sites 1RS, 1BS, 6BS



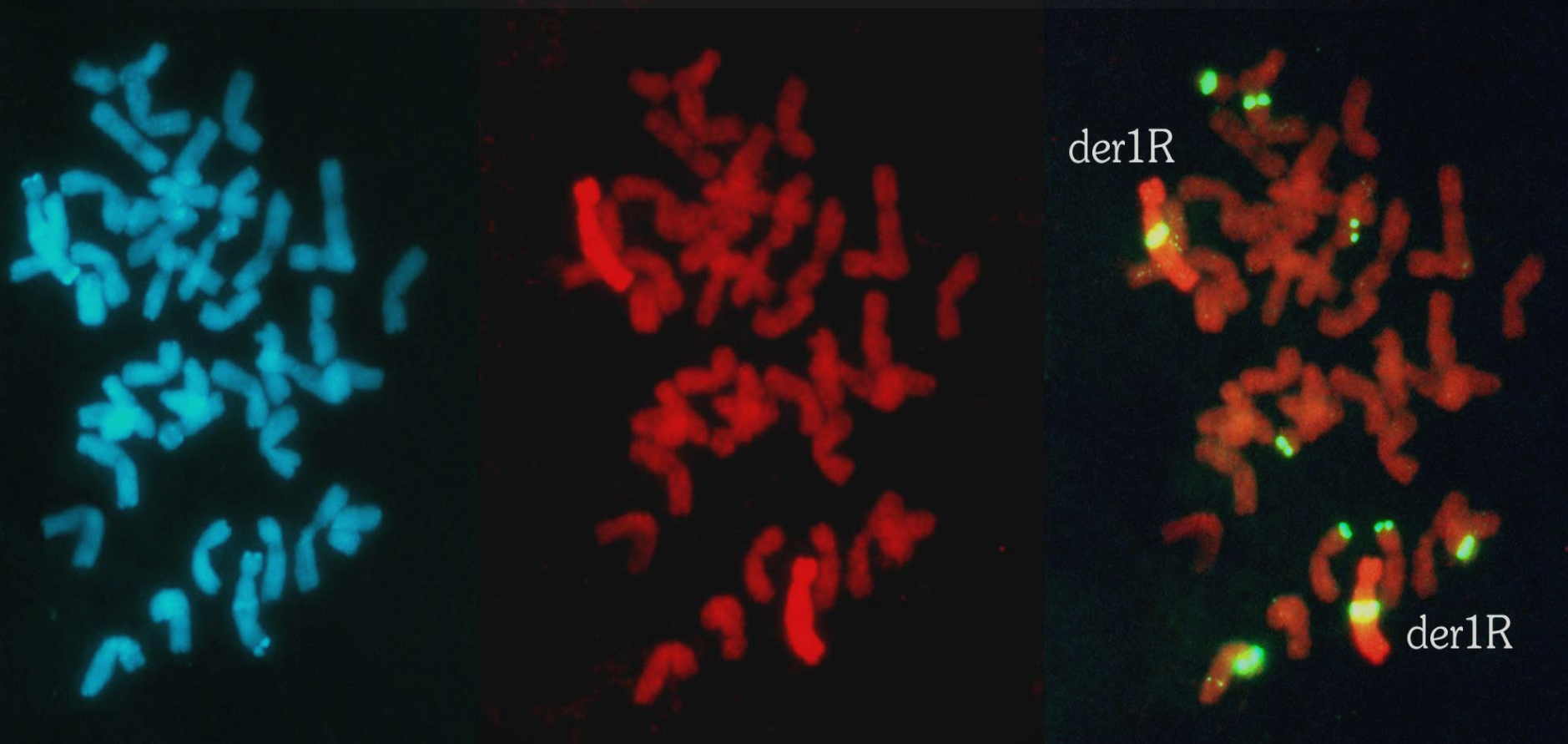
1DL.1RS





# LINE 7-102 DERIVED FROM A TRITICALE X WHEAT CROSS

rye chromosome derivative 1R substitutes wheat chromosome 1D



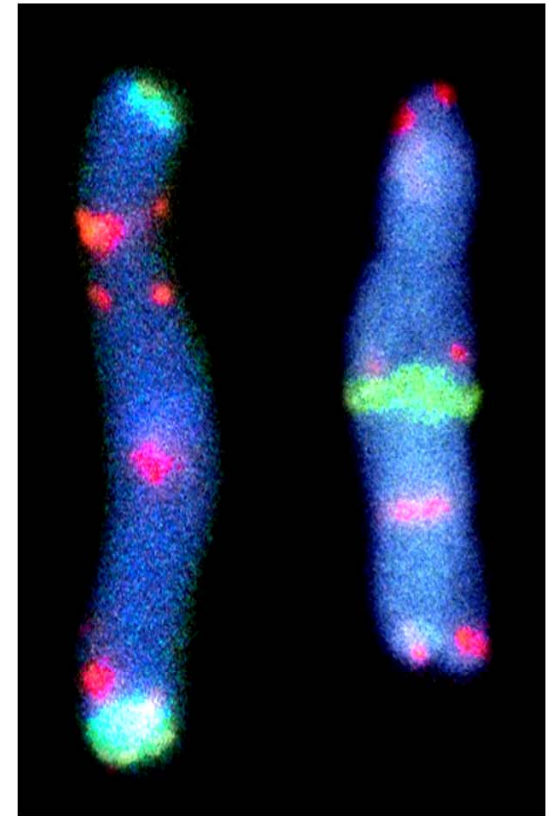
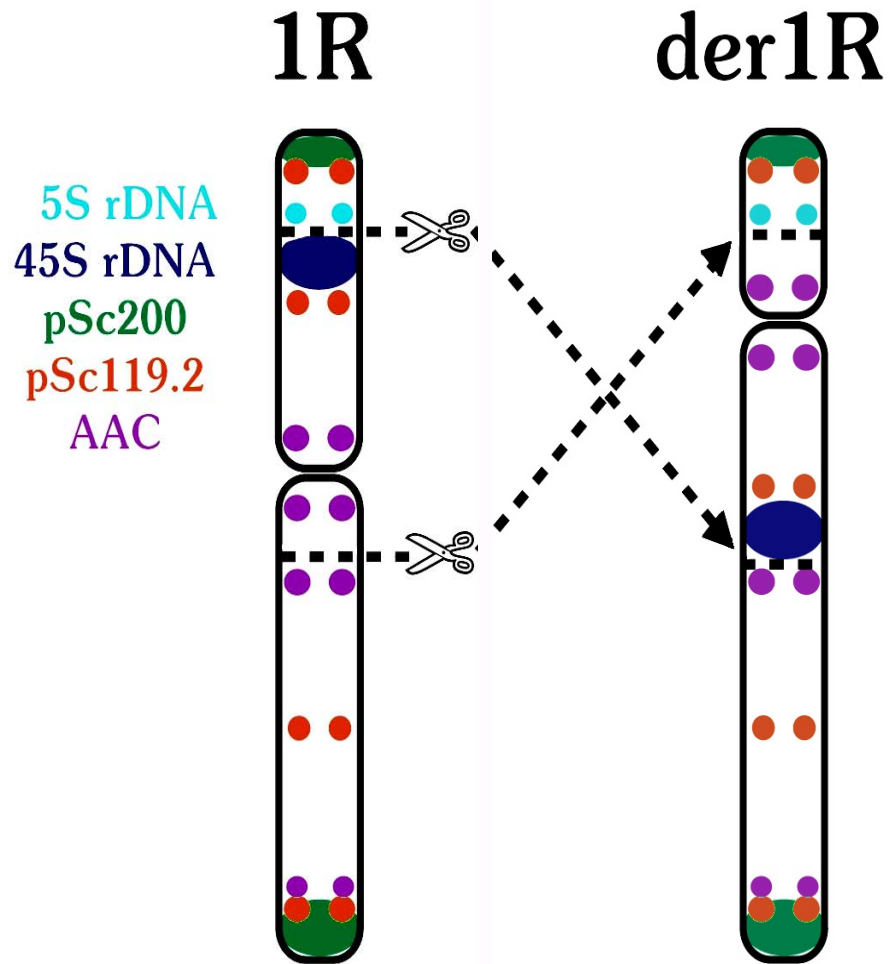
DAPI

Rye genomic DNA probe

pTa71 (45s rDNA probe)



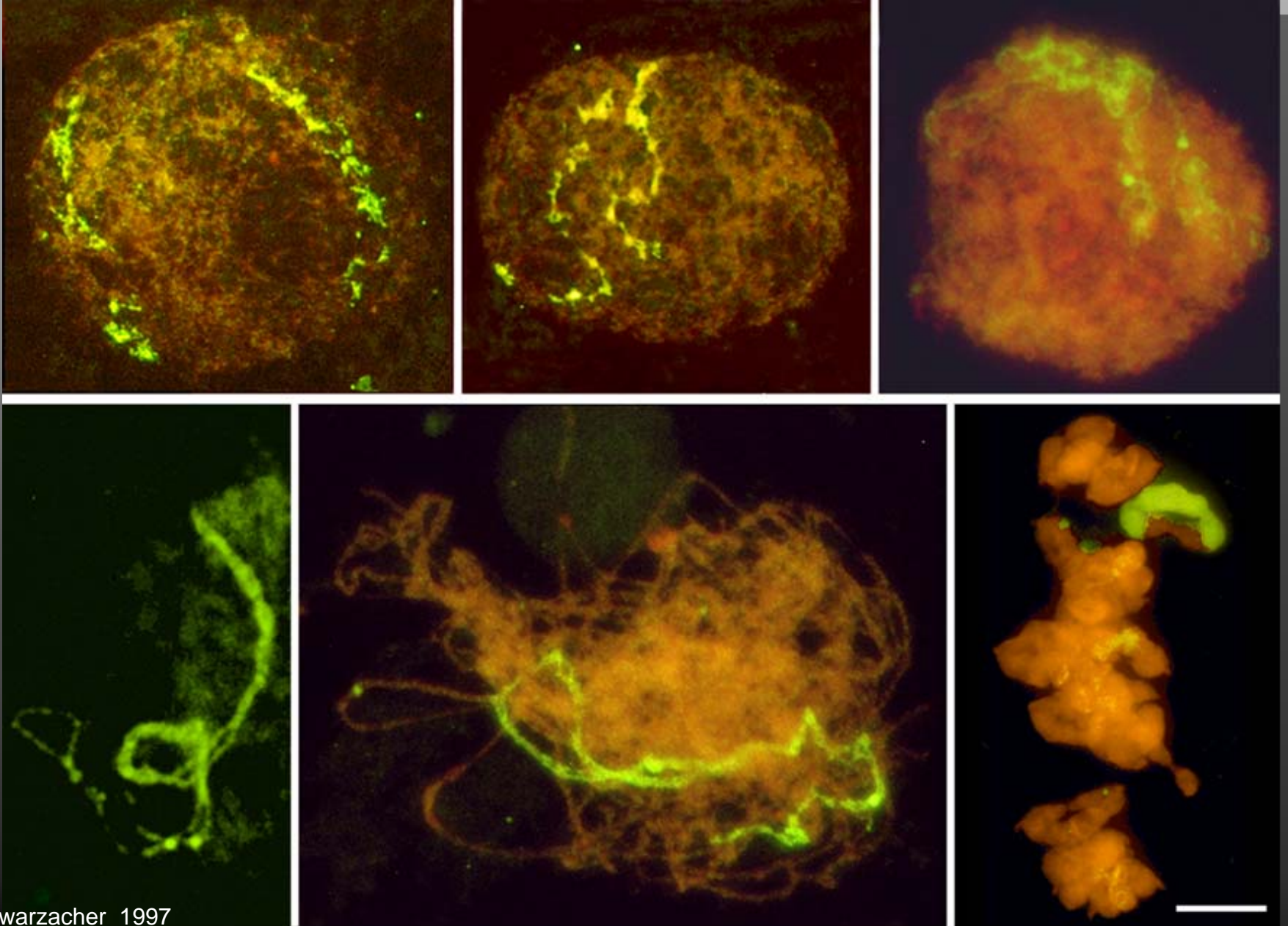
# Derivative chromosome 1R of Lines 7-102 and 7-169



AAC  
pSc200

pSc119.2  
pTa71

# Wheat 5AS.5RL at meiosis



# Characterization of new sources of Wheat streak mosaic virus resistance



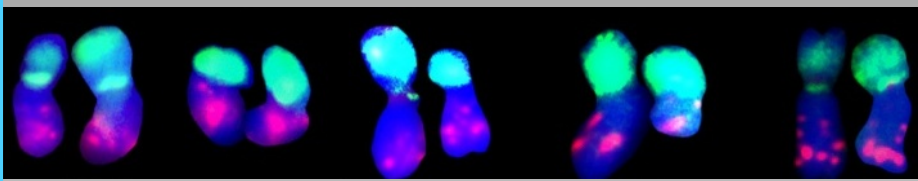
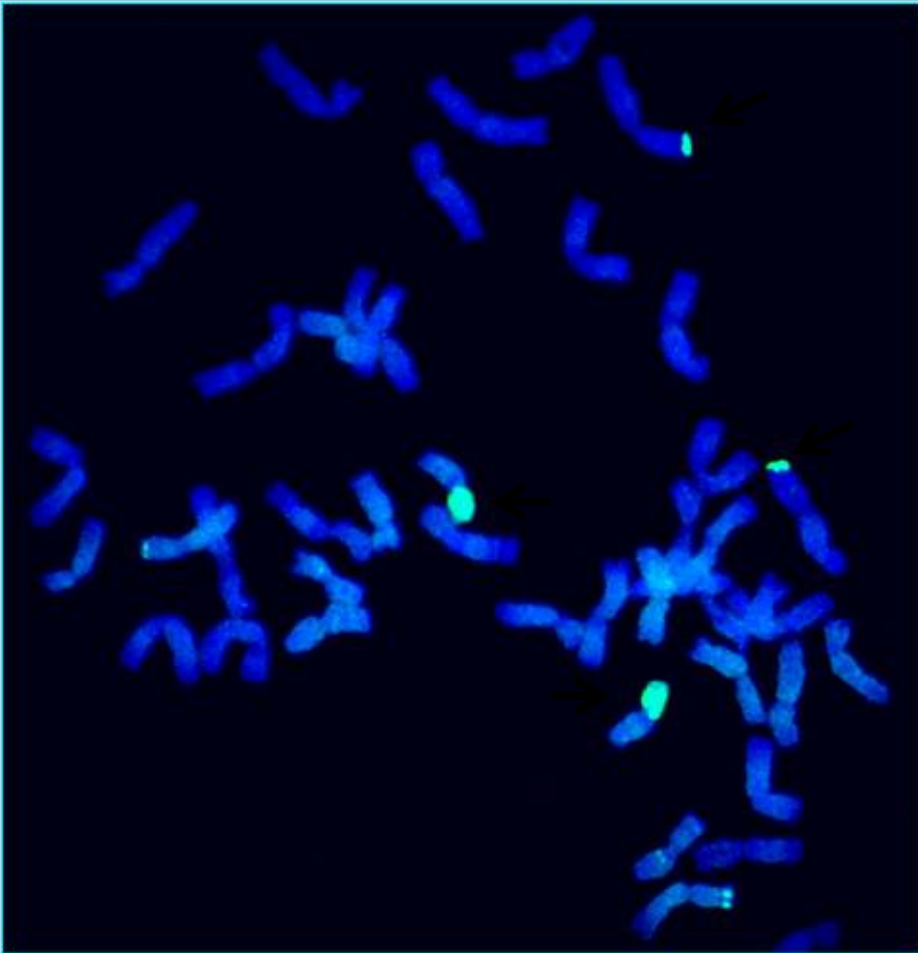
WSMV resistant and susceptible lines in field trials

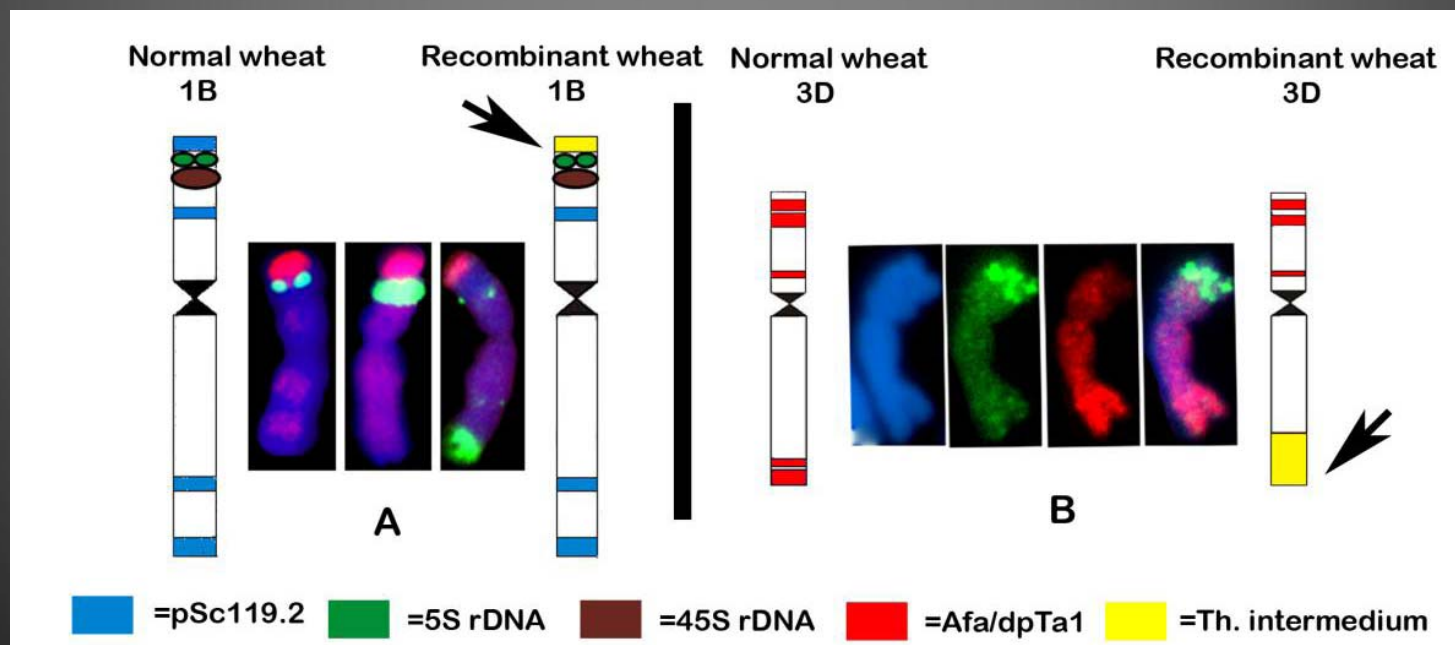
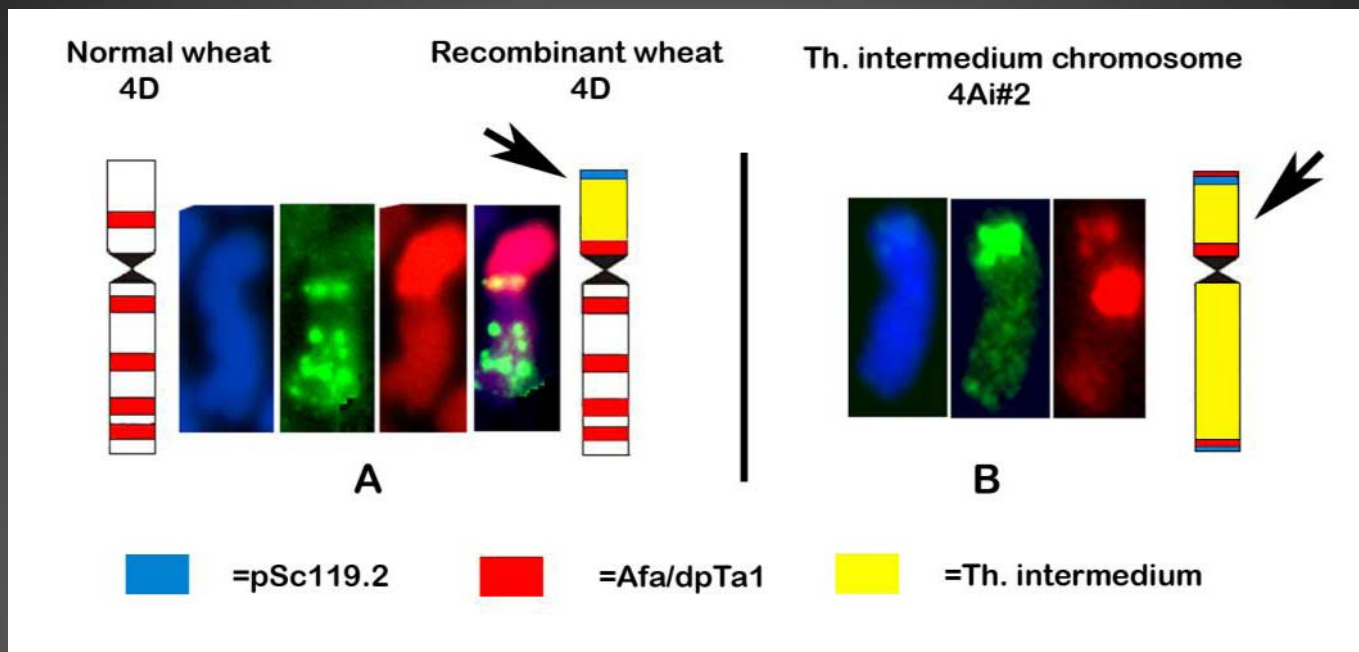
24 resistant and non-resistant sister lines from six different populations of wheat lines that potentially include an alien chromosome arm from *Thinopyrum intermedium* carrying WSMV resistance (Wsm-1 gene)

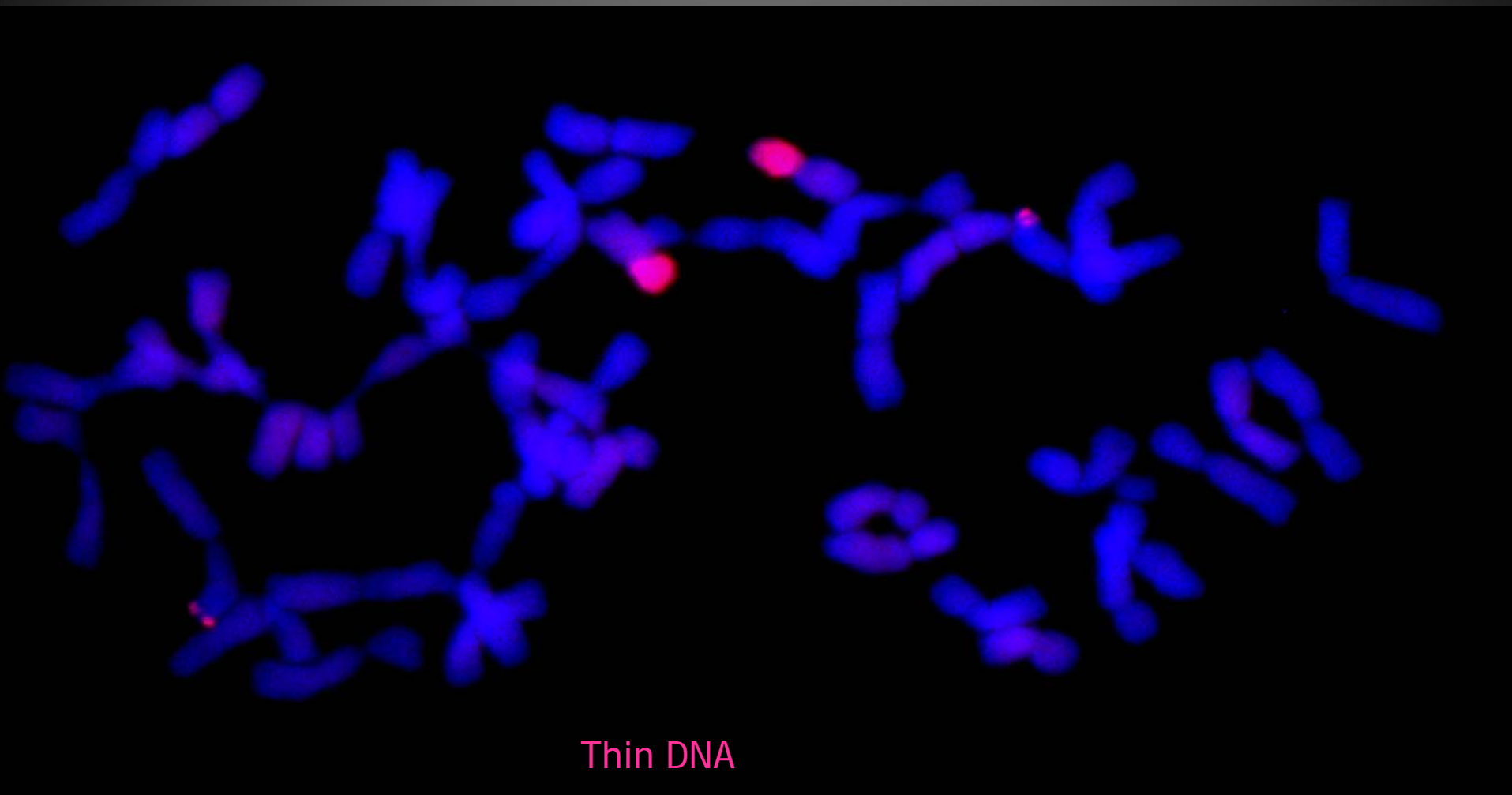
Bob Graybosch, USDA-ARS, University of Nebraska, USA



# WHEAT WITH T4DL.4J<sup>SS</sup> TRANSLOCATION:



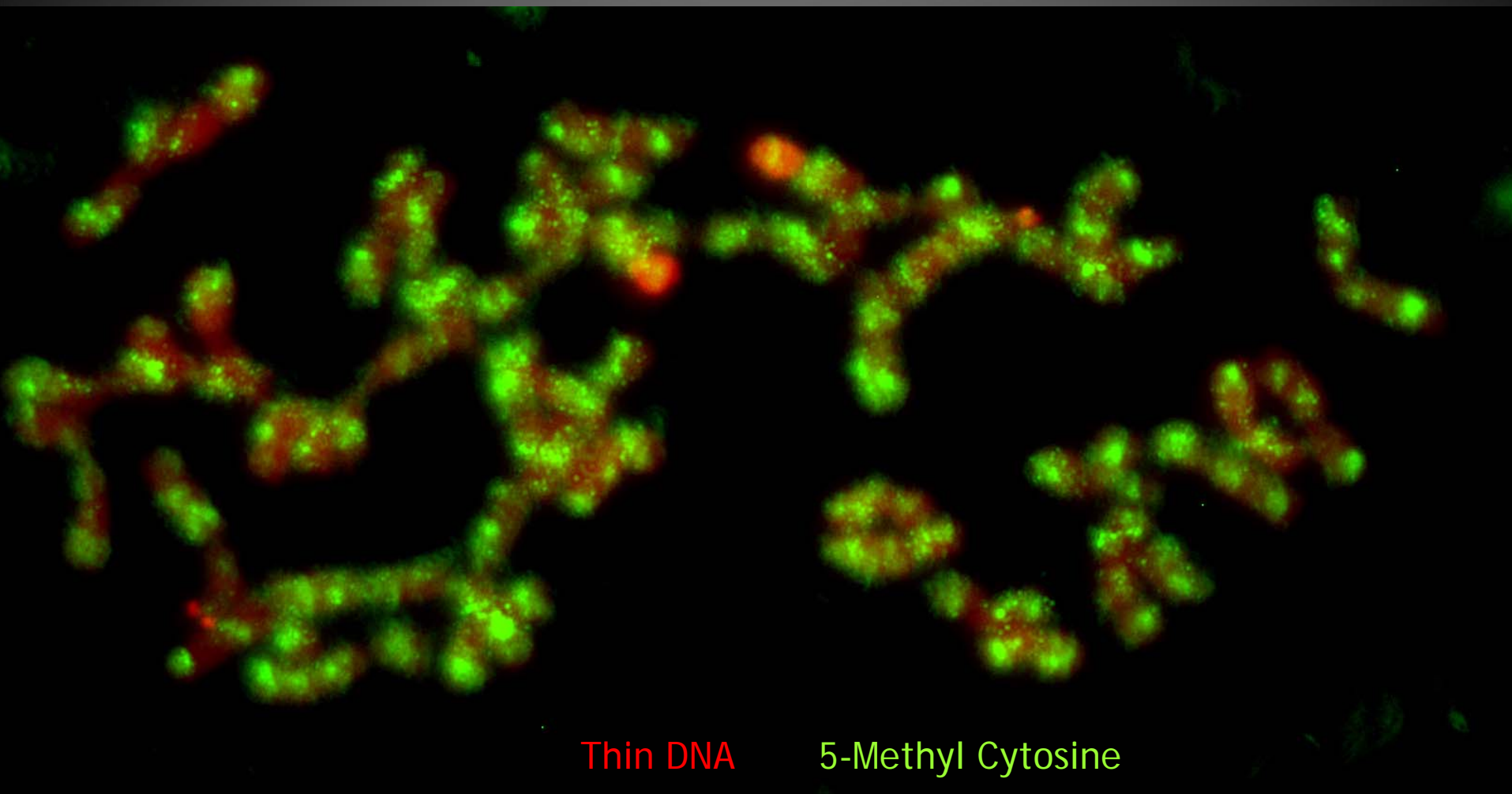




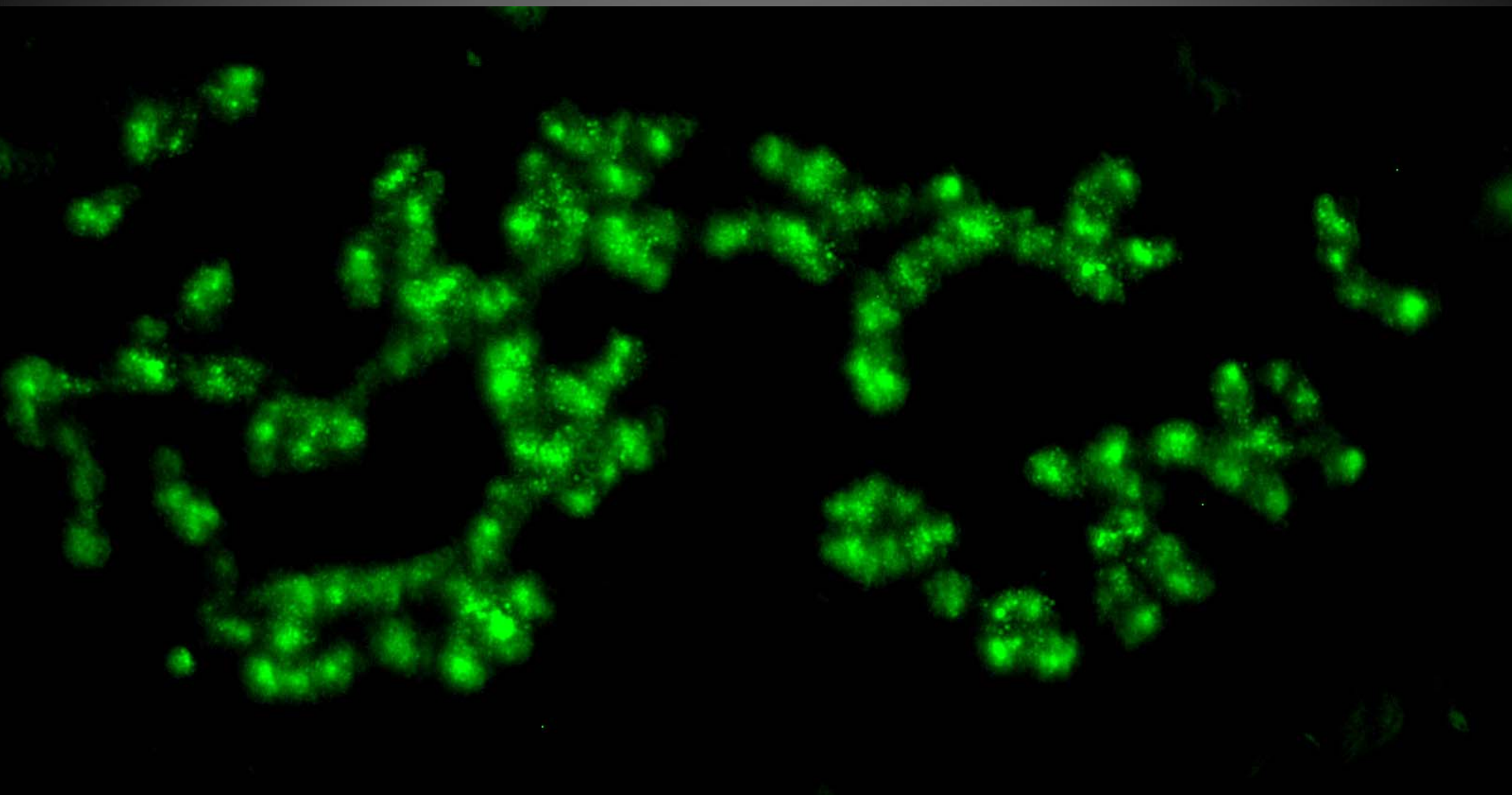
Thin DNA



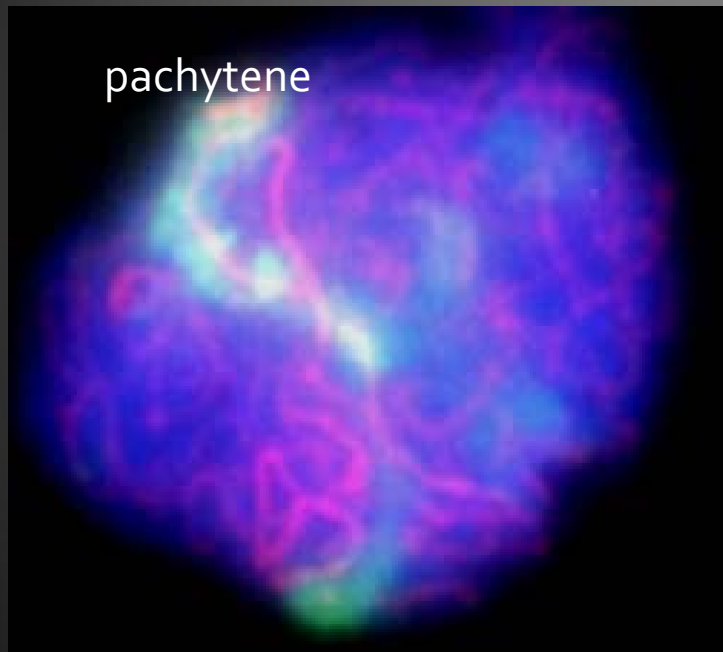
The alien Thin fragment is not heavily methylated



The alien Thin fragment is not heavily methylated



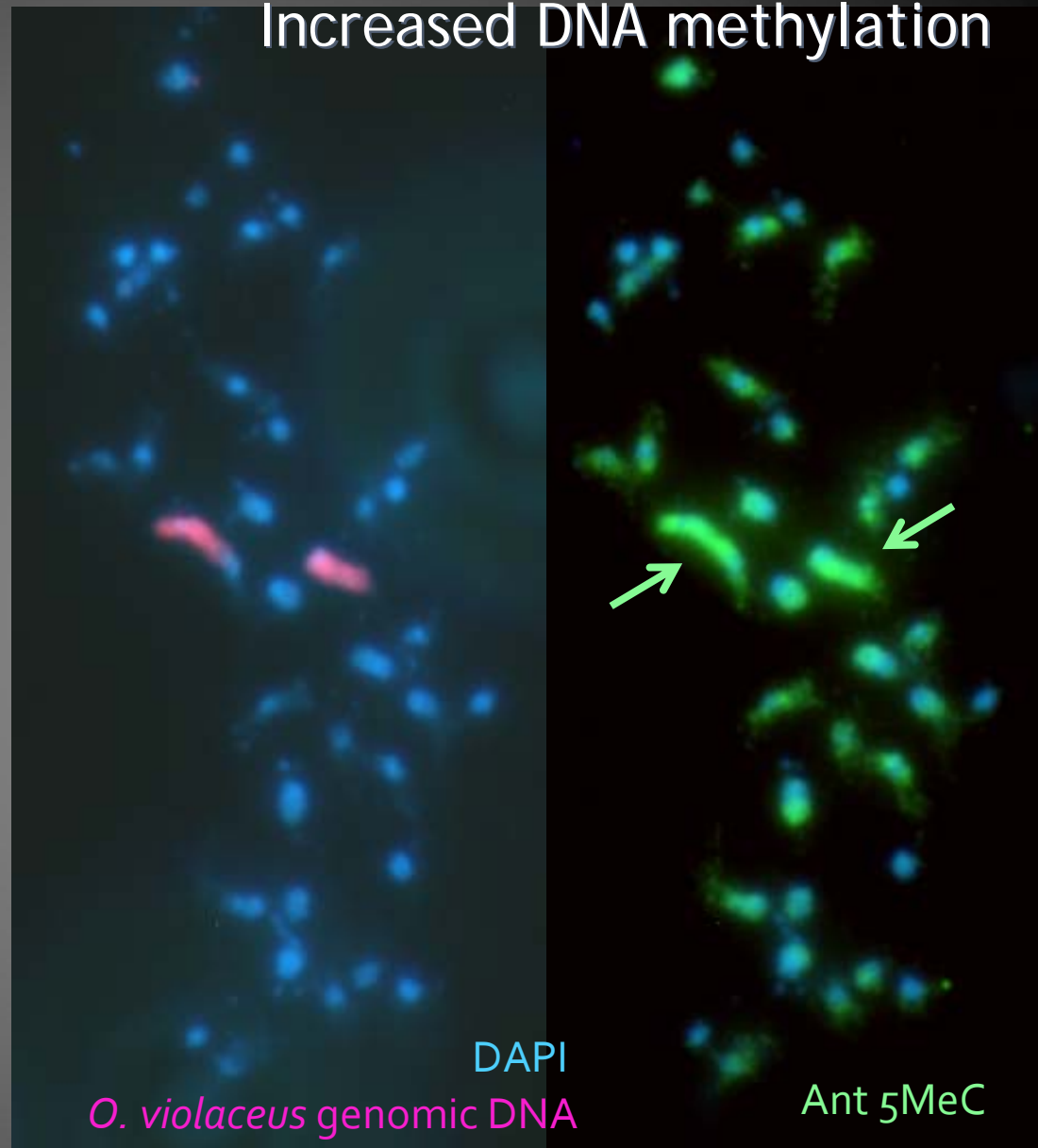
# ADDITION LINE *ORYCHOPHRAGMUS* *VIOLACEUS*



DAPI, ASY-1

*O. violaceus* genomic DNA

Homozygous partly unpaired and partly  
paired with *Brassica* chromosomes



Increased DNA methylation

DAPI

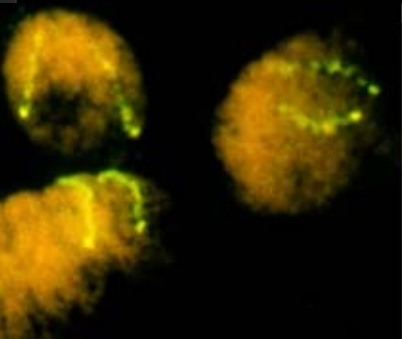
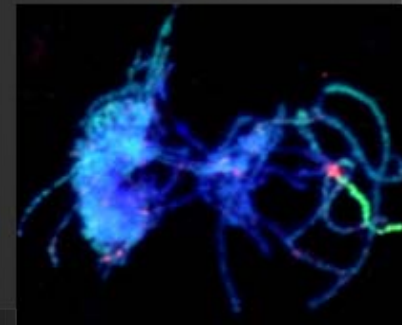
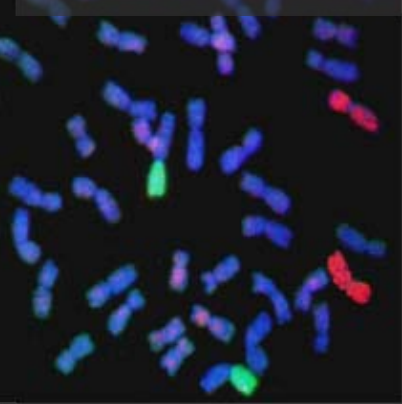
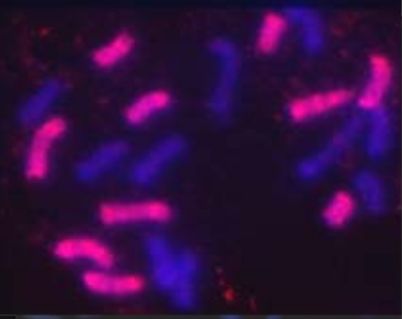
*O. violaceus* genomic DNA

Ant 5MeC



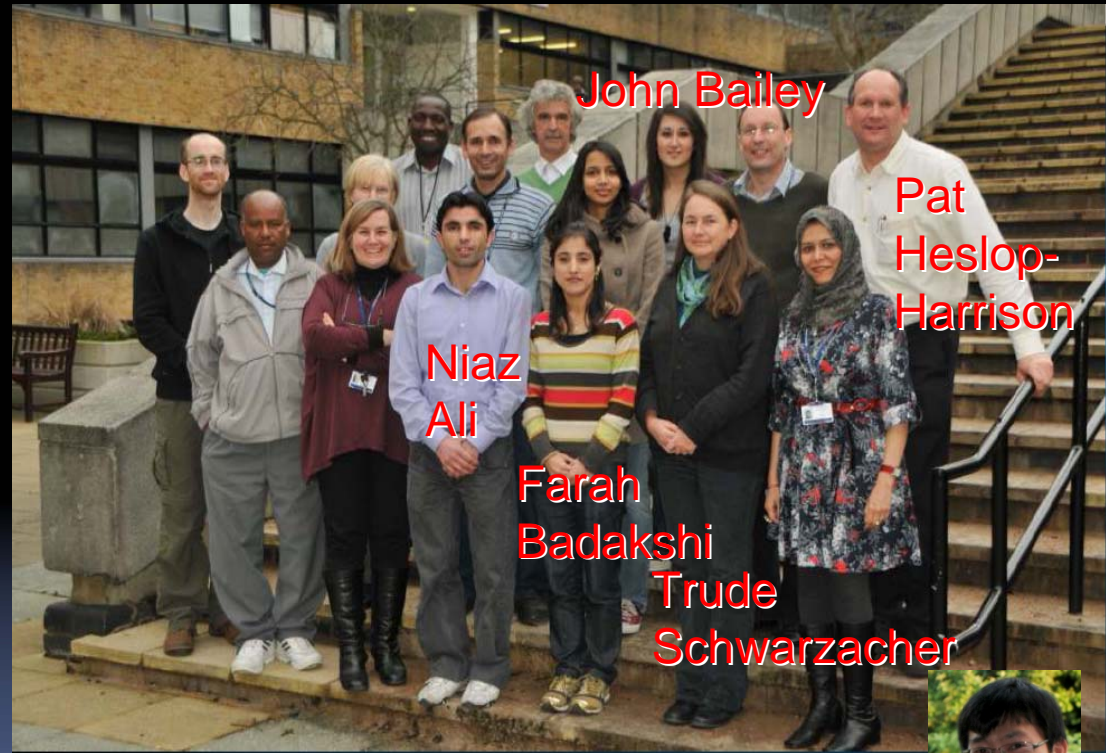
# GENOMIC *IN SITU* HYBRIDIZATION GISH

- Parental origin of hybrids
  - Auto or allo-poliploidy
- Alien chromatin in breeding lines
  - Size and origin
  - Recipient chromosome
- Meiosis and chromosome pairing
- Interphase cytogenetics
- Understanding hybrid genomes
  - Chromosome behaviour
  - Chromatin function



# Thank you to members of the Molecular cytogenetics lab

- Collaborators and past members of the lab
  - Richard Gornall, Leicester
  - Bob Graybosch, Nebraska, USA
  - Karine Alix, INRA/Univ Paris-Sud, France



ANNALS OF  
BOTANY  
AoBBlog.com

[www.molcyt.com](http://www.molcyt.com)



EU-Crocusbank

Xianhong Ge ,  
Huazhong Agricultural  
University, Wuhan,  
China

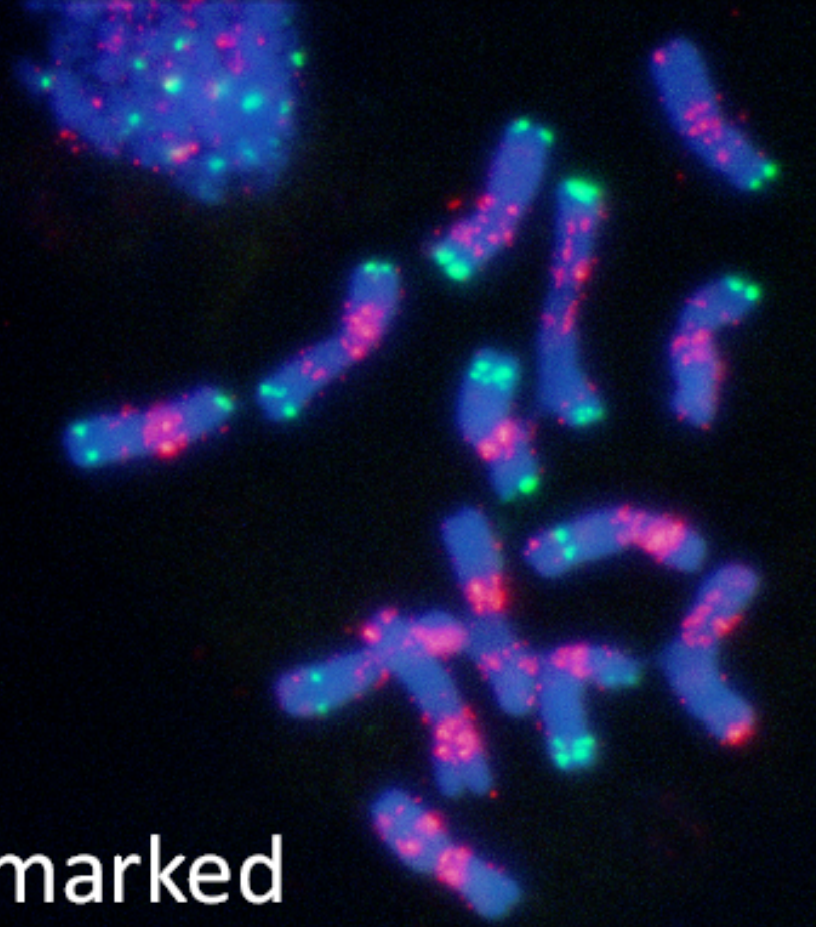




The Origin of Species begins with an example of how humans have domesticated certain species and used artificial selection, in contrast to natural selection, to produce extraordinary variation in a short time. Darwin focused on “fancy” pigeons, but it is apparent in many domestic species, including dogs, cattle and crop plants like wheat (see below).

At that time, it was not known how such variation arose or was maintained. Now we know that this variation is due to genes and chromosomes, and a team here in Leicester investigate how the number and organisation of chromosomes varies across wheat varieties.

Dr Trude Schwarzacher and Prof Pat Heslop-Harrison investigate the evolution of cereals by examining their chromosomes. On the right are the chromosomes from a root cell of rye. The red and green spots are DNA sequences which can be used to trace the ancestry of different cereal species and varieties.



“Species are only strongly marked  
and permanent varieties”

Below are examples of wheat ears from different hybrids and wild varieties of wheat.

