

# RNA-mediated Transgenerational Epigenetic Inheritance of DNA Rearrangements and Copy Number

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Plant and Animal  
Genome XX  
San Diego, CA  
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# *Oxytricha trifallax*

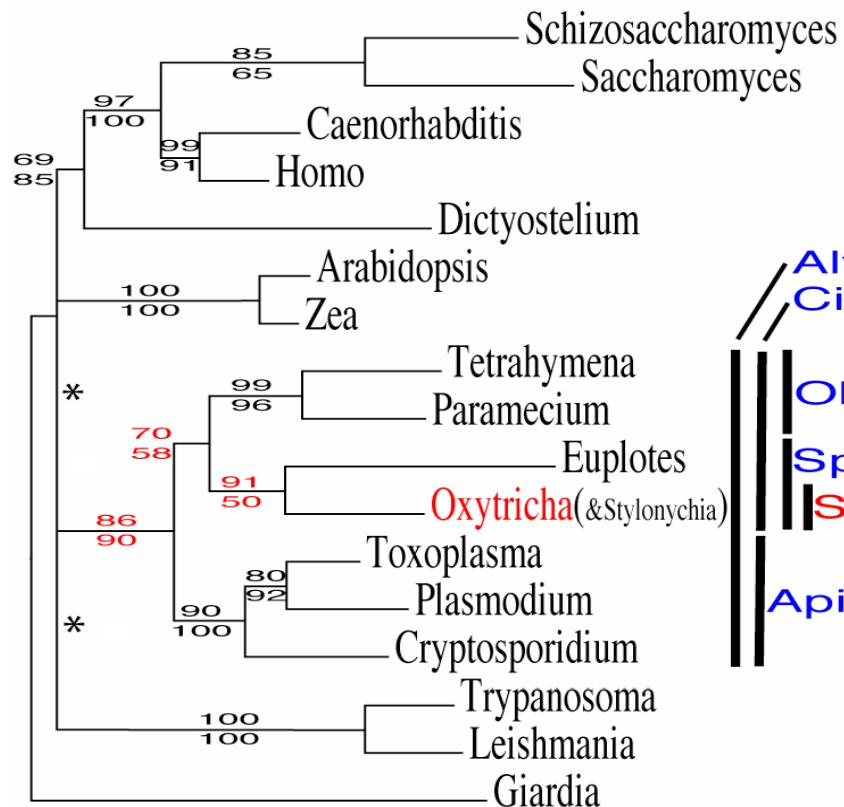


Fig. 1  
50 steps  
\* low-value nodes  
removed from original



Modified from Baldauf et al. (2000)

# *Oxytricha trifallax*

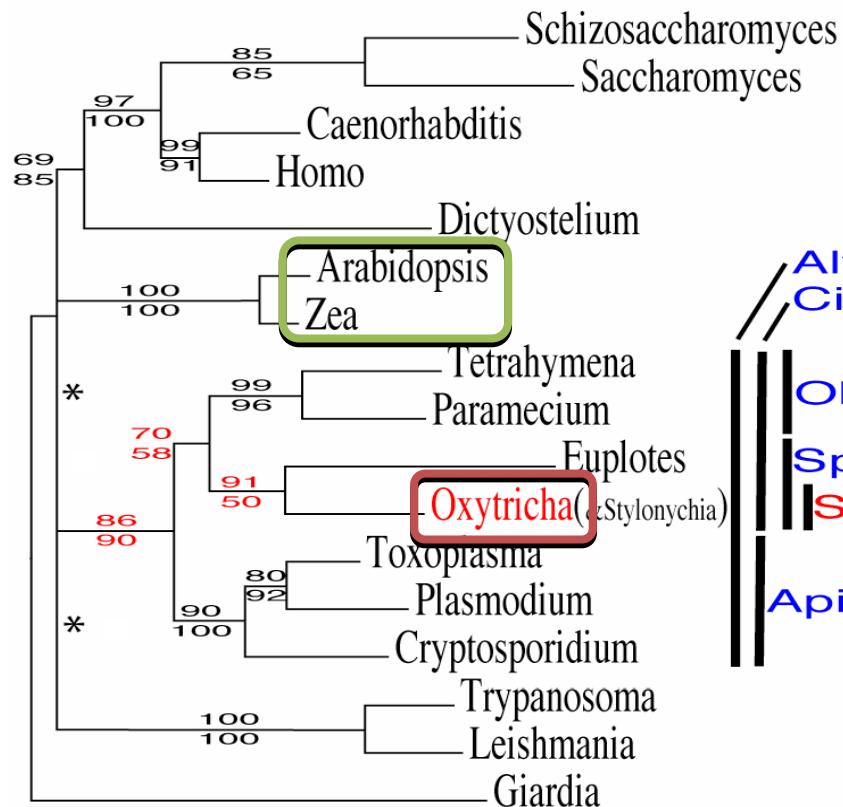


Fig. 1  
50 steps  
\* low-value nodes  
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Alveolate  
Ciliophora  
Oligohymenophorea  
Spirotrichea  
Stichotrichia  
Apicomplexa



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# *Oxytricha trifallax*

*Oxytricha* to *Homo* represents approximately 1.8 billion years of independent evolutionary history

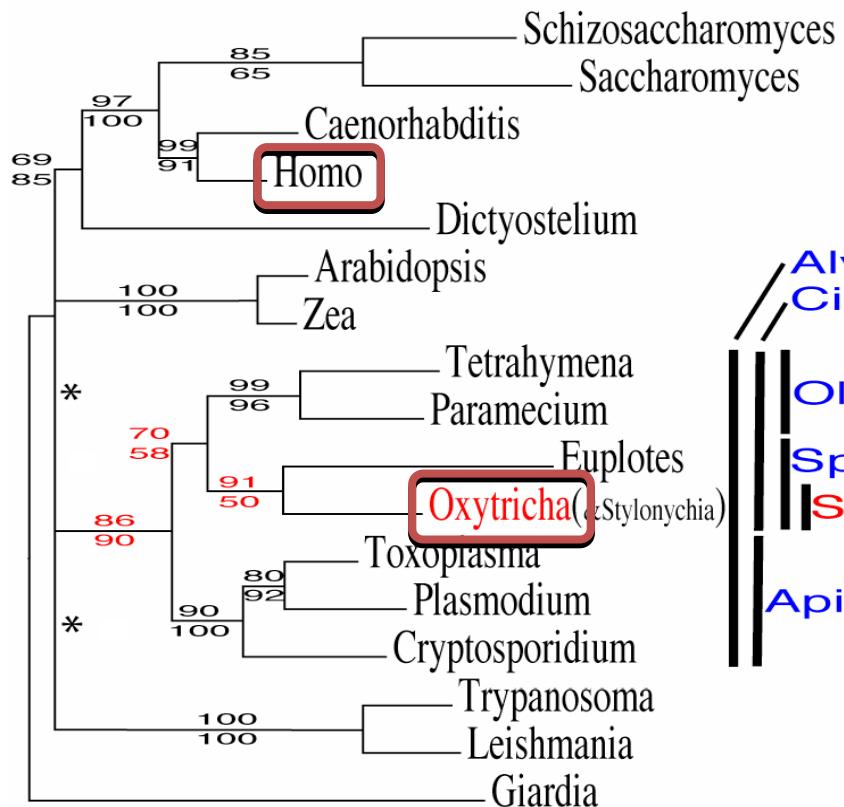


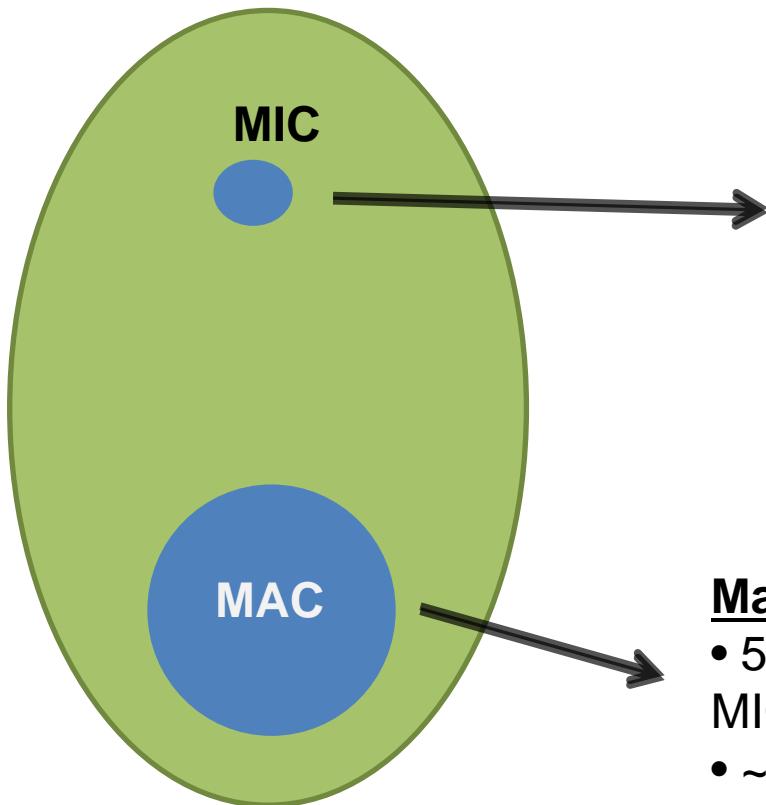
Fig. 1  
50 steps  
\* low-value nodes removed from original



Nei M, Xu P, Glazko G.  
PNAS. 2001 Feb 27;98(5):2497-502.

Modified from Baldauf et al. (2000)

# Oxytricha Nuclear Dimorphism



Oxytricha cell

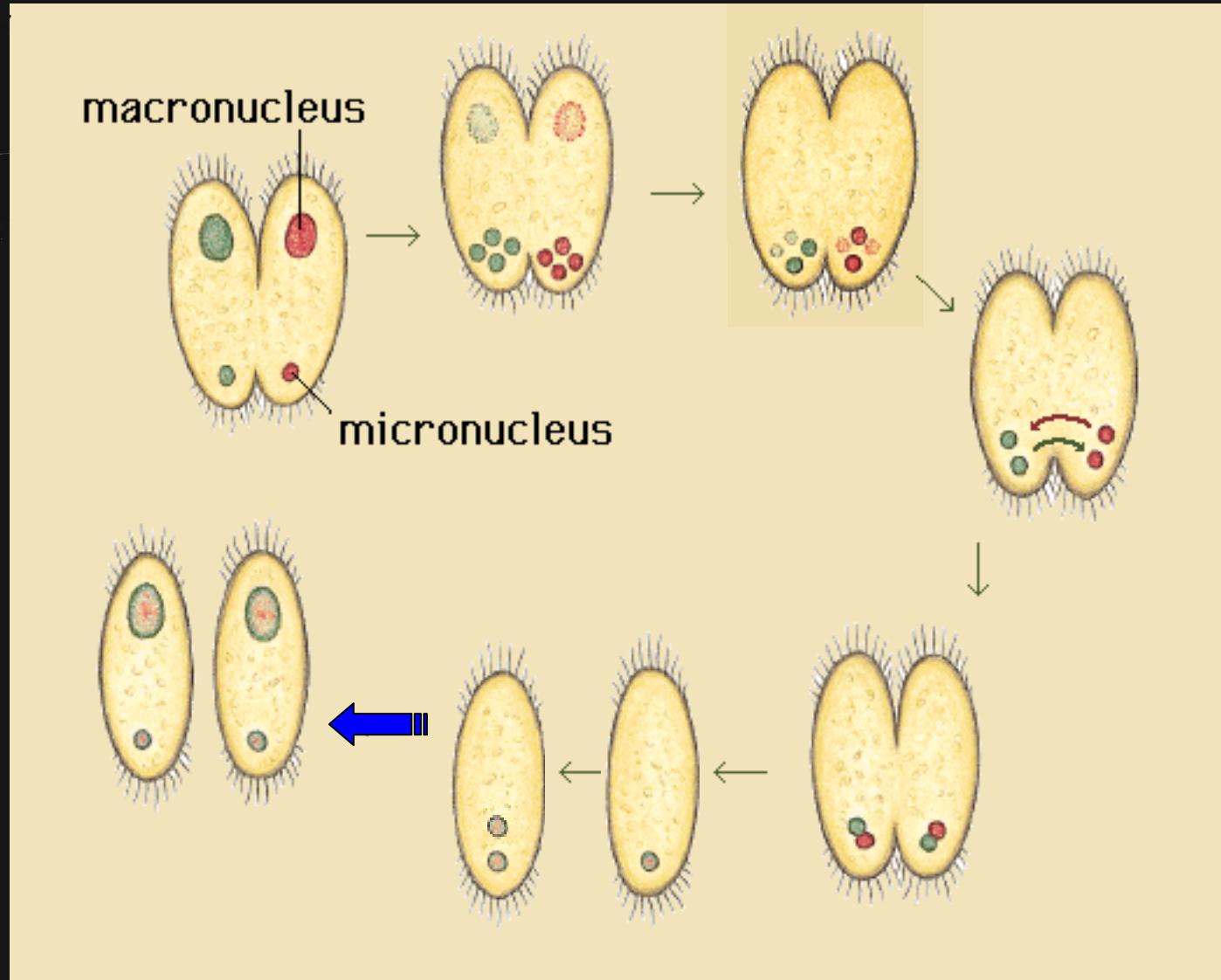
## Micronucleus:

- 1 gigabase genome ( $1 \times 10^9$  bp)
- 120 euchromosomes
- diploid
- scrambled genes
- direct repeats, transposons
- transcriptionally silent
- “dark matter”

## Macronucleus:

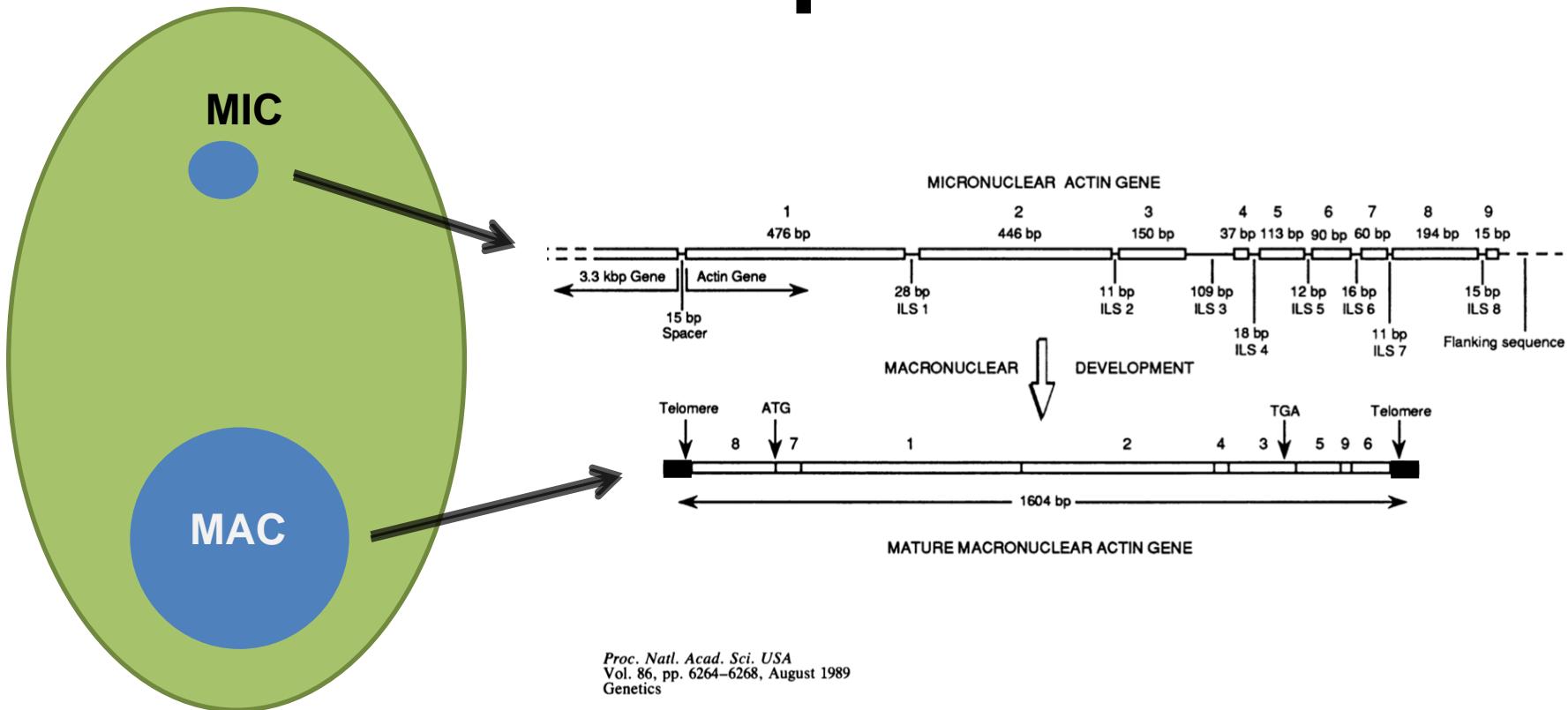
- 50 megabase genome ( $50 \times 10^6$  bp) (20x smaller than MIC)
- ~20,000 gene-sized nanochromosomes
- nanochromosomes average 3.2kb long
- Usually 1 gene per nanochromosome
- 1900n
- Short telomeres
- No centromeres
- Protein expression

# The Oxytricha Sexual Cycle



SEM courtesy Bob Hammersmith

# Oxytricha Nuclear Dimorphism



Oxytricha cell

Reordering of nine exons is necessary to form a functional actin gene in *Oxytricha nova*

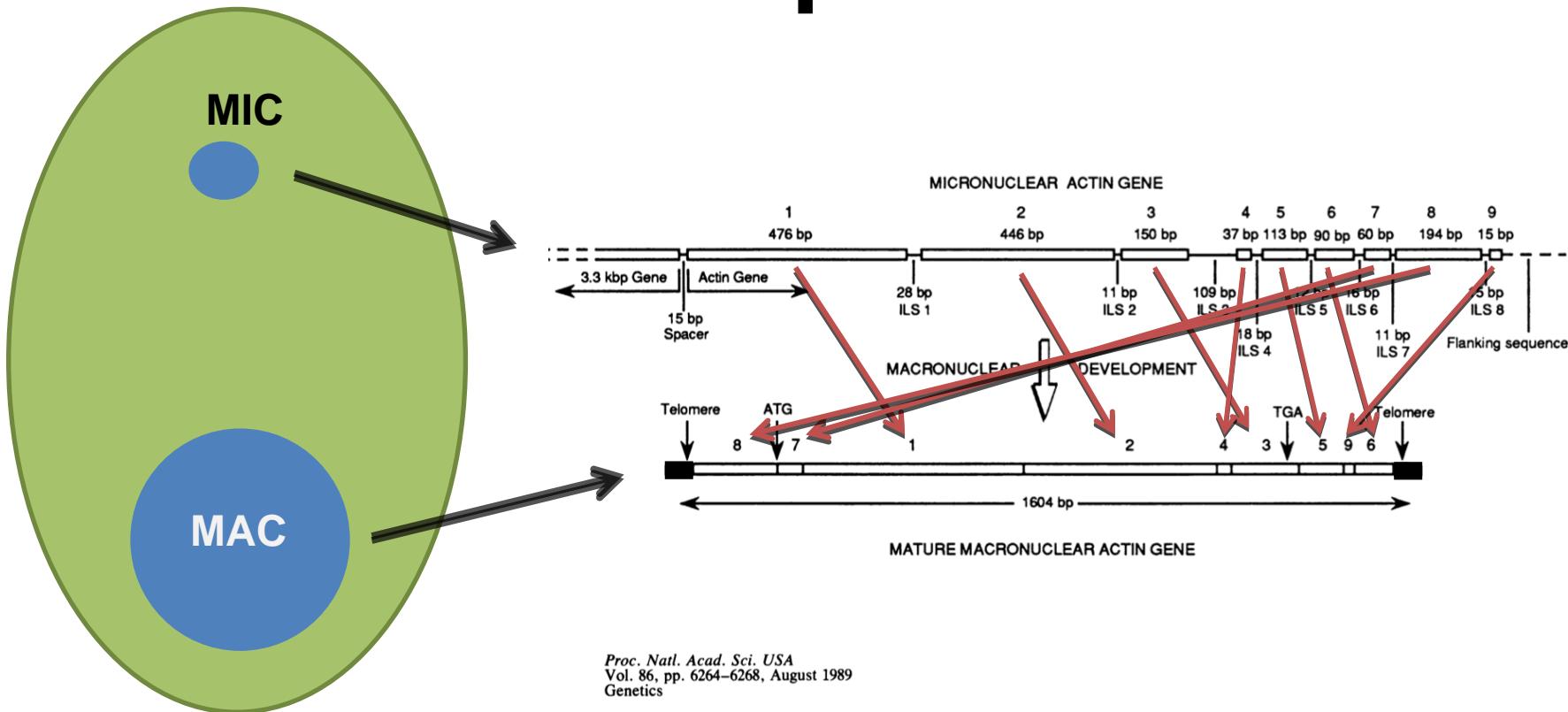
(macronuclear development/micronucleus/hypotrichs)

ARTHUR F. GRESLIN, DAVID M. PRESCOTT, YOSHIO OKA\*, STEPHEN H. LOUKIN†, AND JAMES C. CHAPPELL

Department of Molecular, Cellular, and Developmental Biology, University of Colorado, Boulder, CO 80309

Contributed by David M. Prescott, May 30, 1989

# Oxytricha Nuclear Dimorphism



*Proc. Natl. Acad. Sci. USA*  
Vol. 86, pp. 6264–6268, August 1989  
Genetics

**Reordering of nine exons is necessary to form a functional actin gene in *Oxytricha nova***

(macronuclear development/micronucleus/hypotrich)

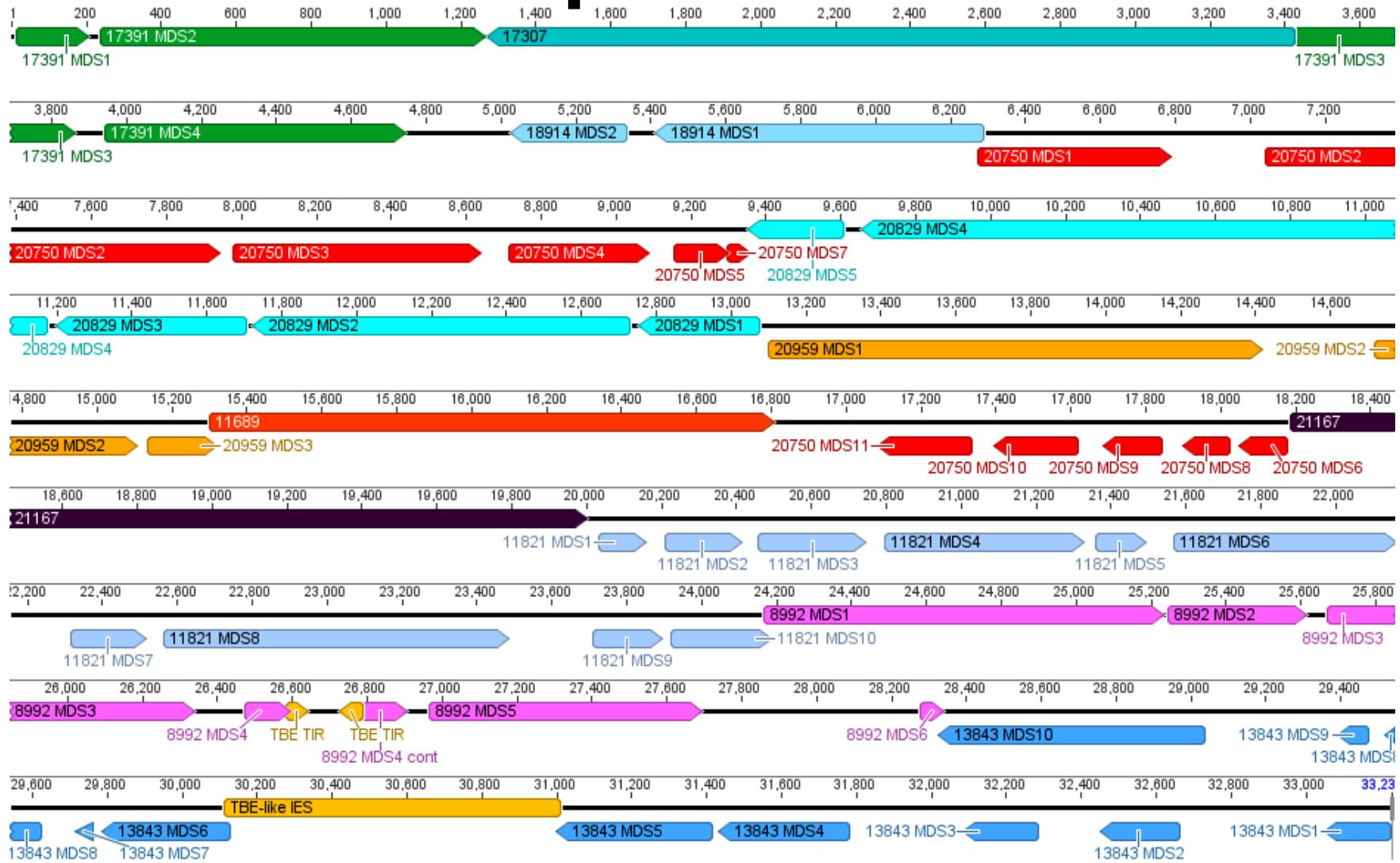
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Oxytricha cell

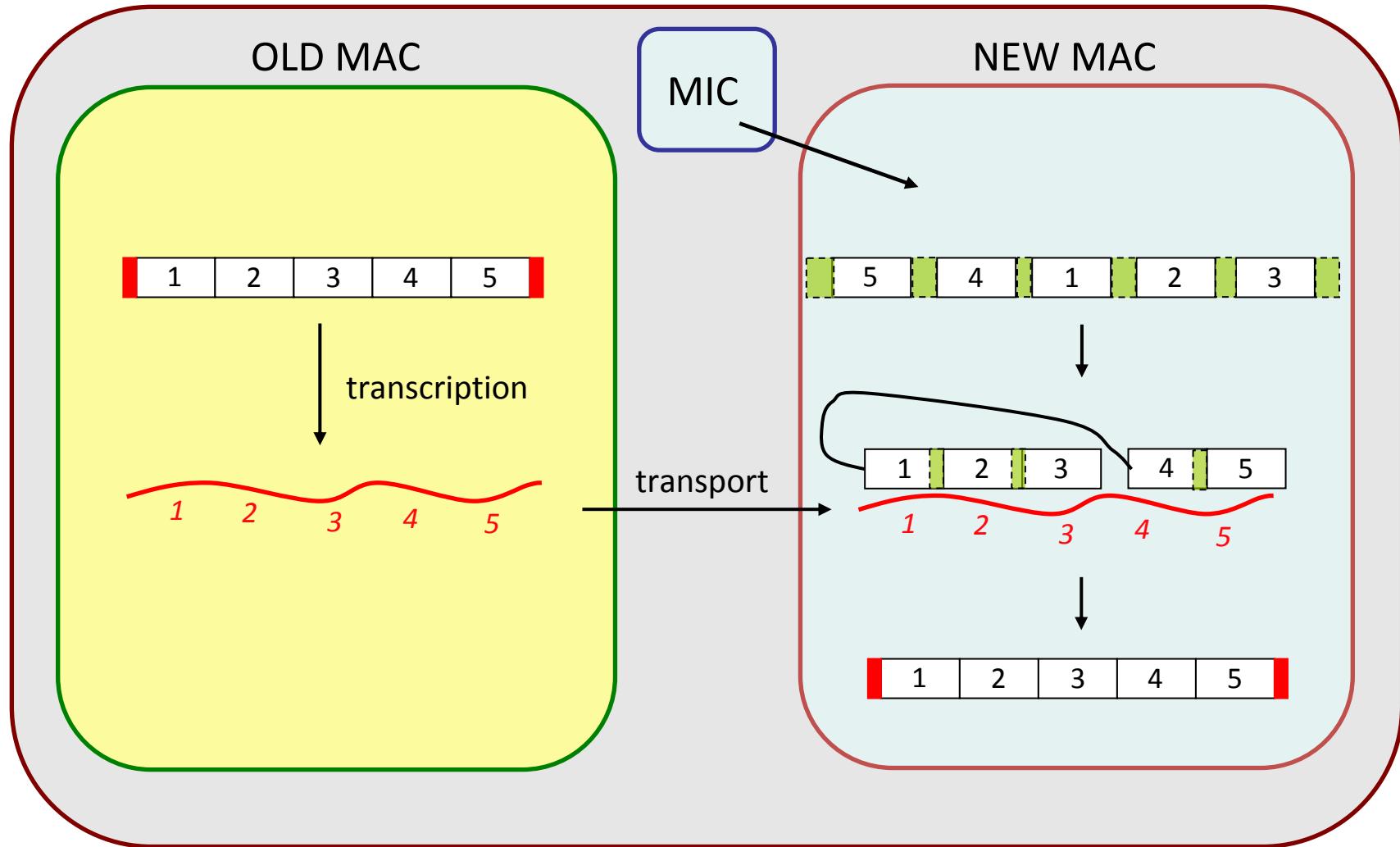
# A MIC mashup of 13 genes in 55 pieces



# Questions

- How does the cell decide to delete or retain a given DNA segment (g  
95%  
deleted!)  
  
? e
- How does the cell know the correct order of pieces scramble genome?
- What machinery performs genome rearrangements?

# Template Model



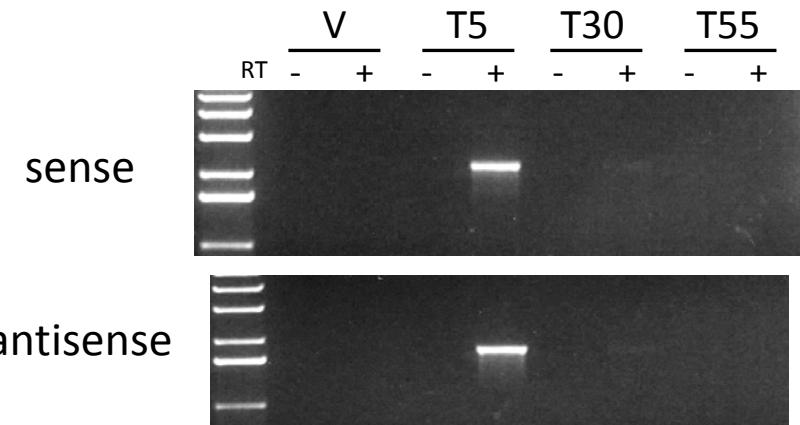
Mariusz Nowacki



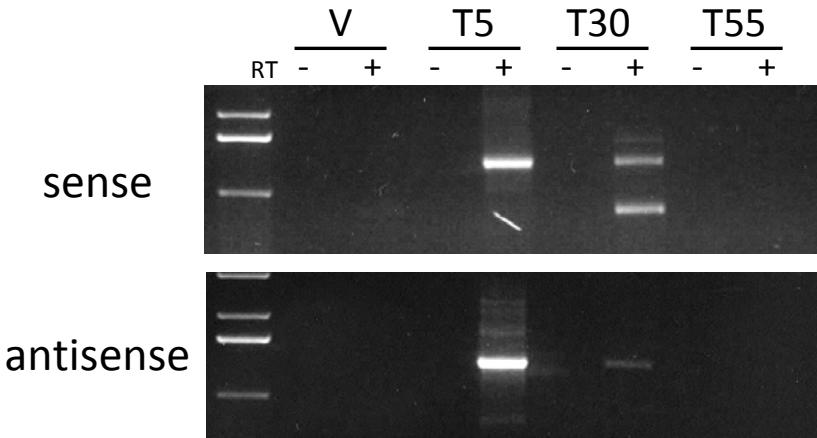
University of Bern

# Telomere RT-PCR can detect putative Template RNAs

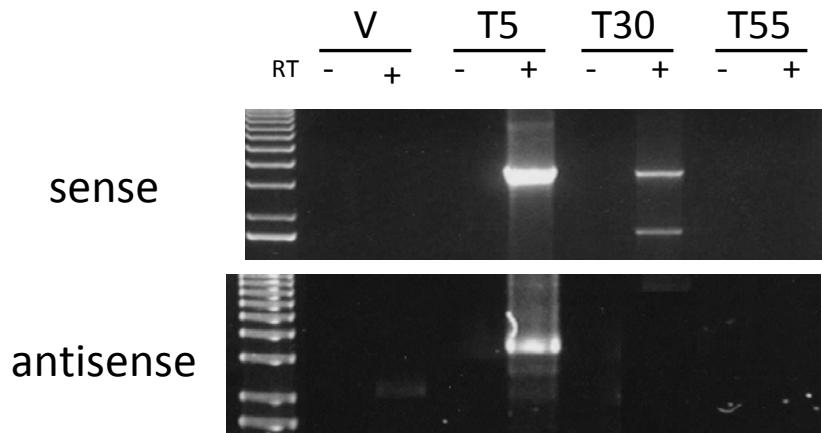
TEBP $\alpha$  (scrambled)



TEBP $\beta$  (non-scrambled)

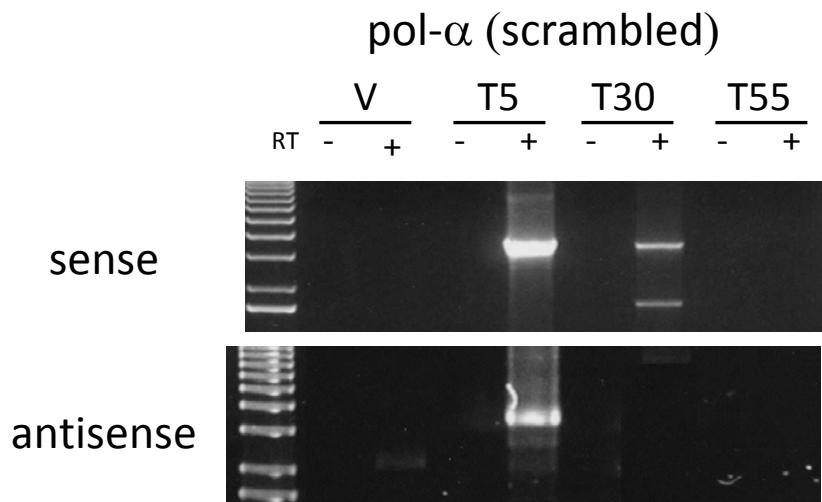
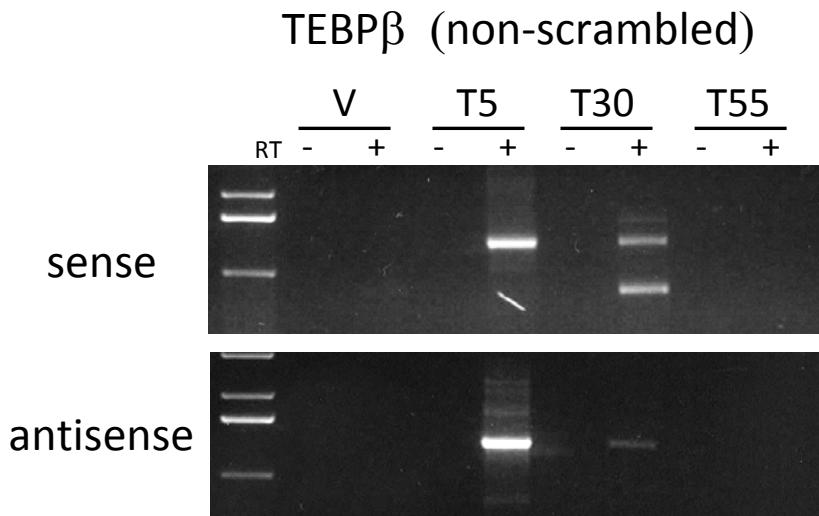
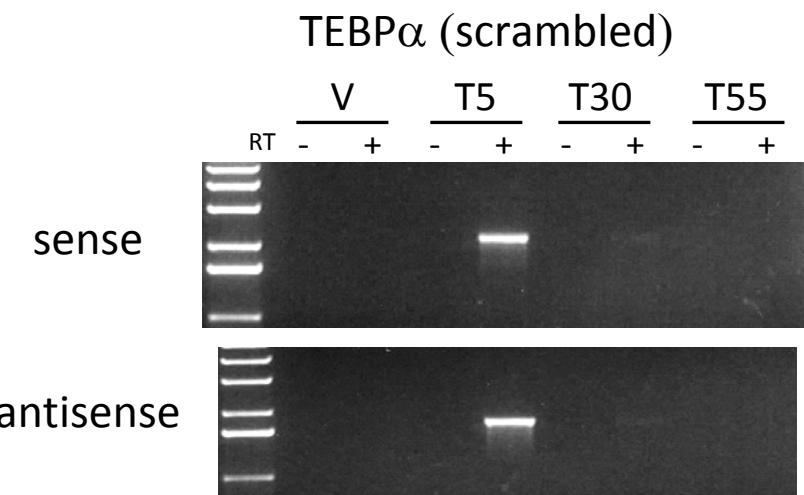


pol- $\alpha$  (scrambled)

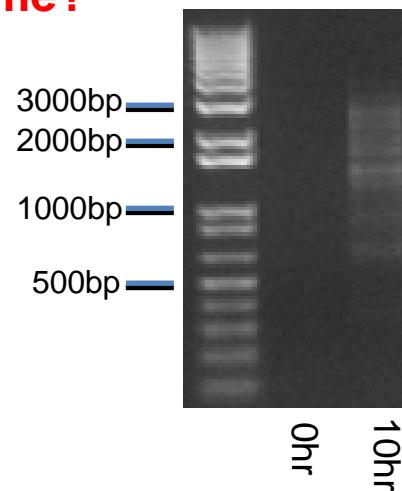


An RNA cache of the entire genome?

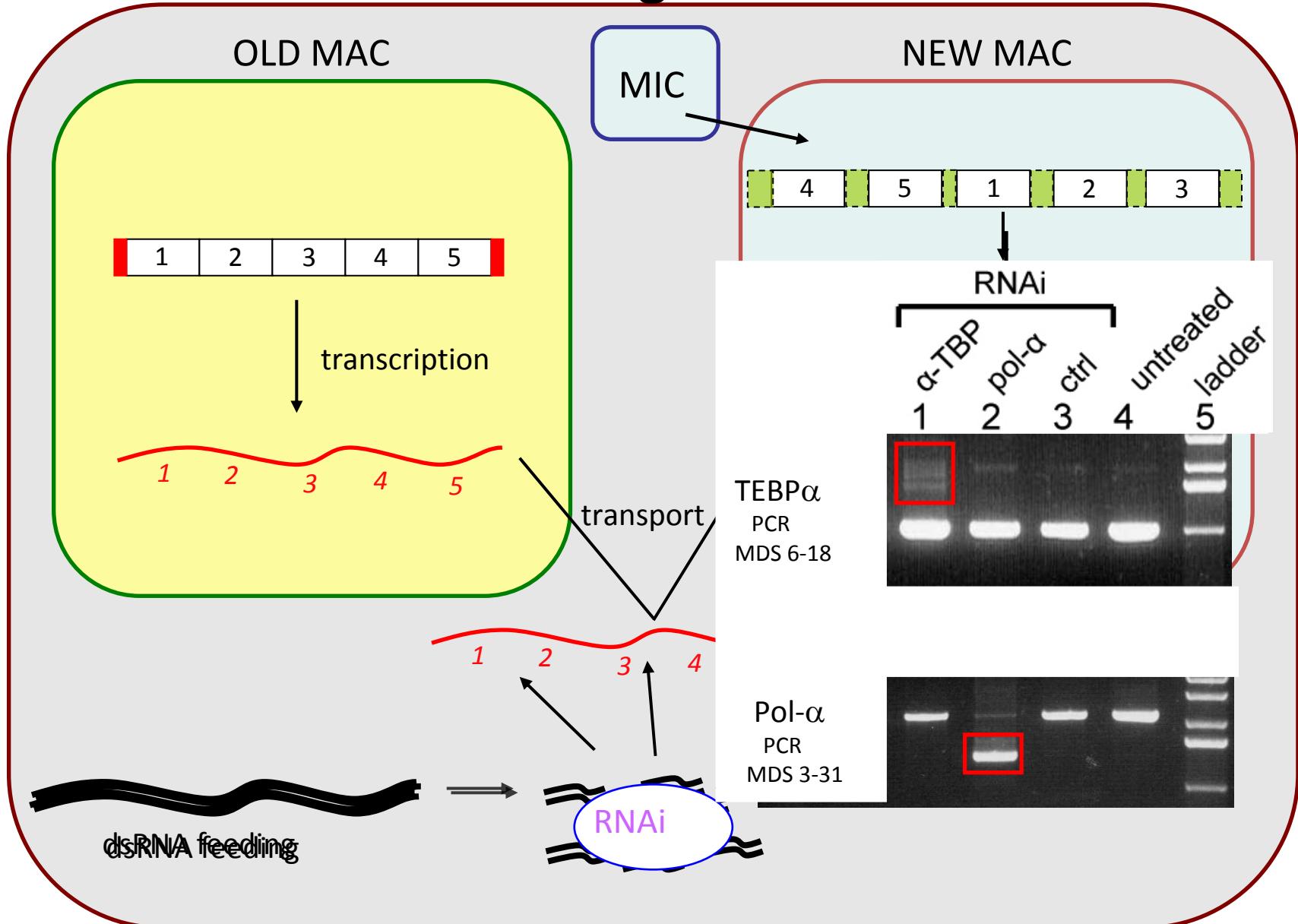
# Telomere RT-PCR can detect putative Template RNAs



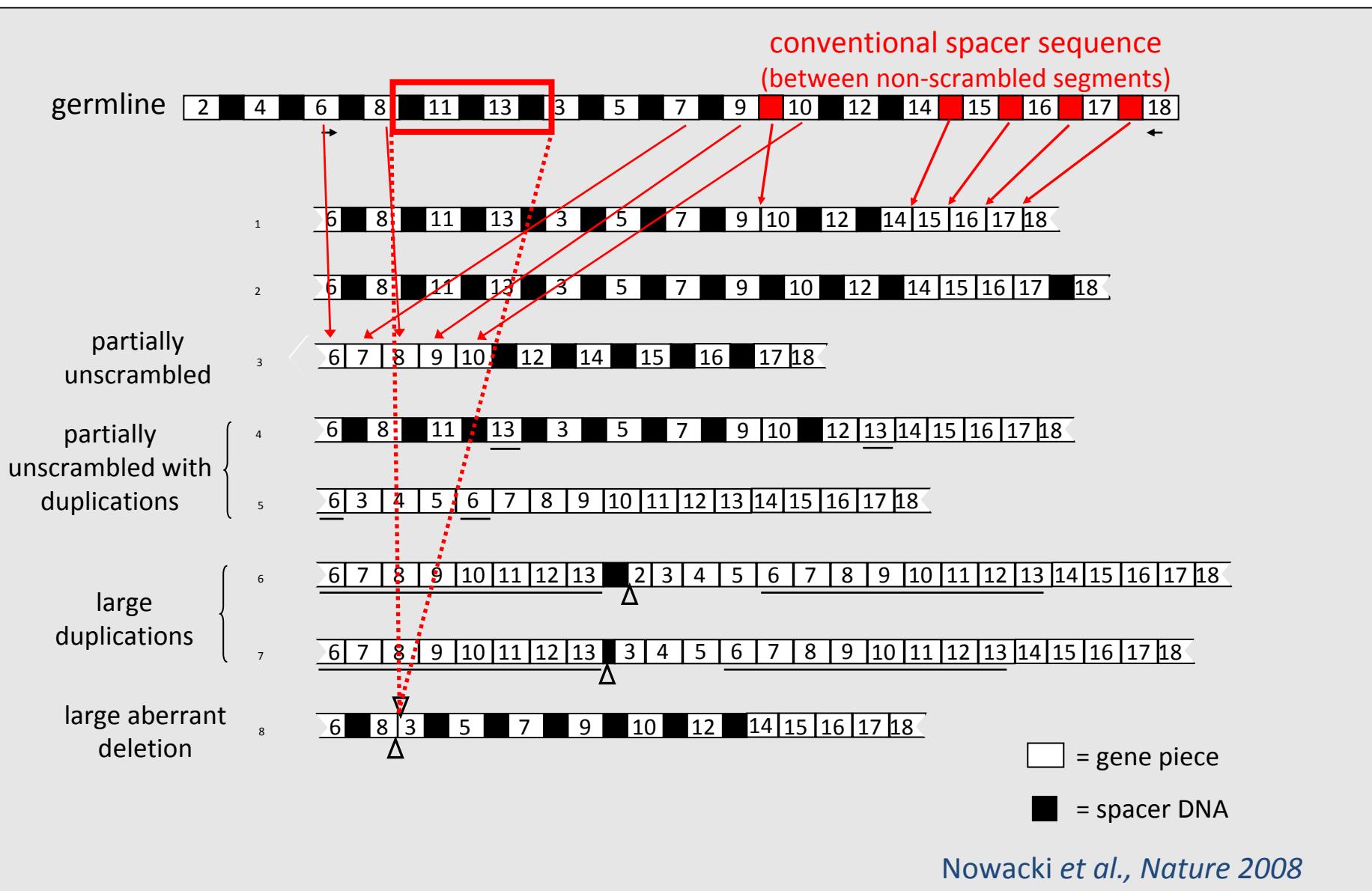
An RNA cache of the entire genome?



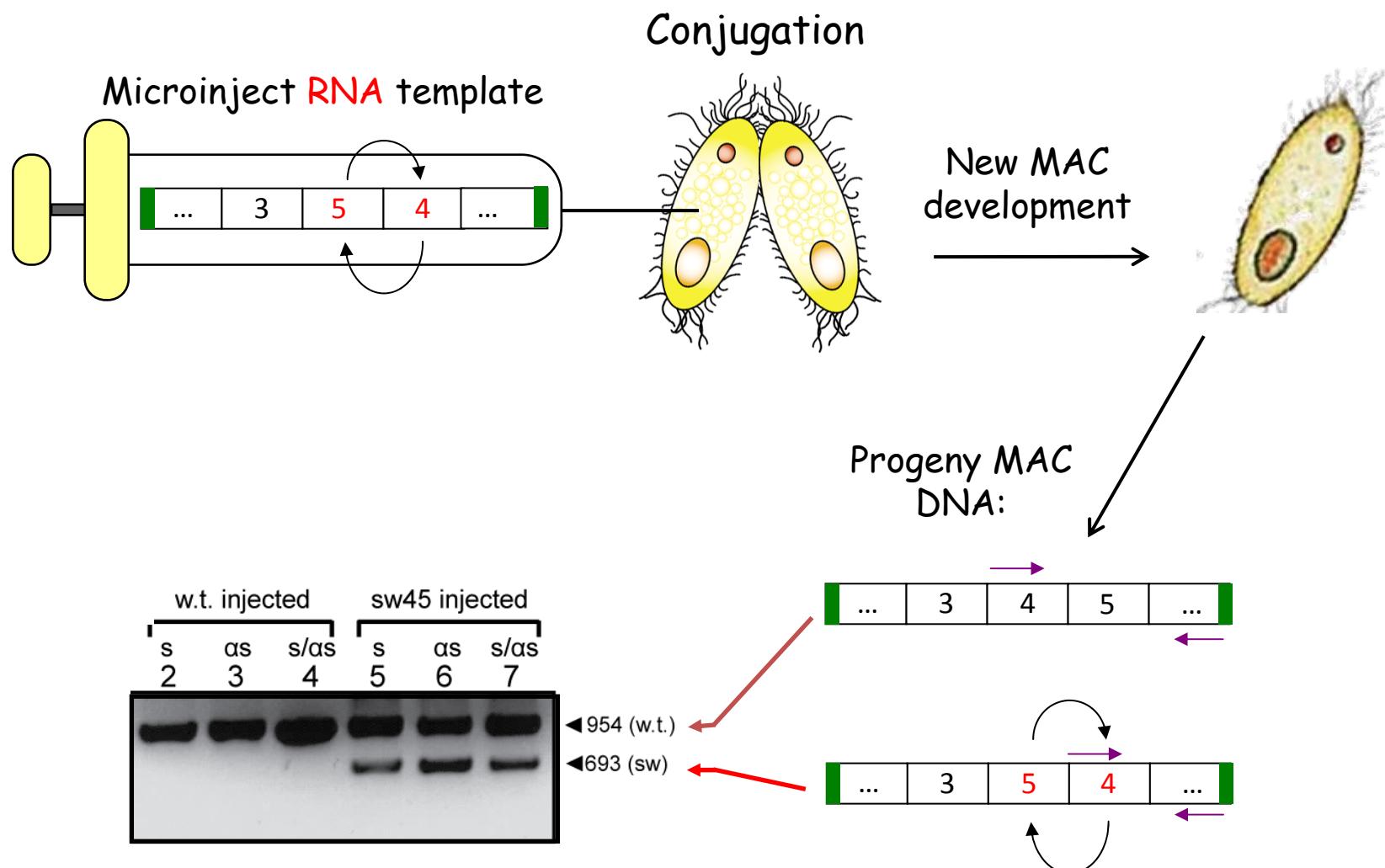
# RNAi against putative templates stalls rearrangements



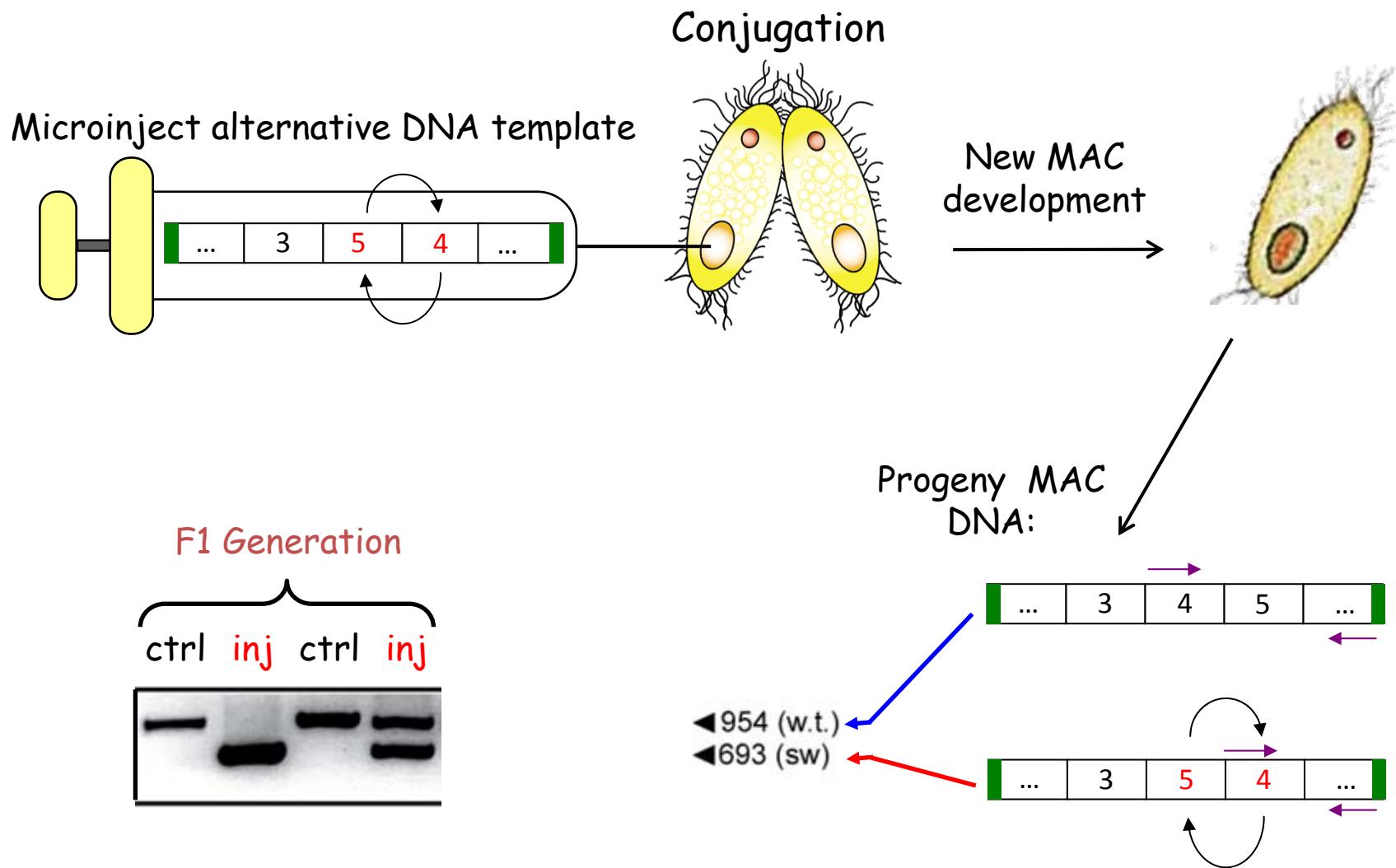
# Examples of aberrantly rearranged molecules



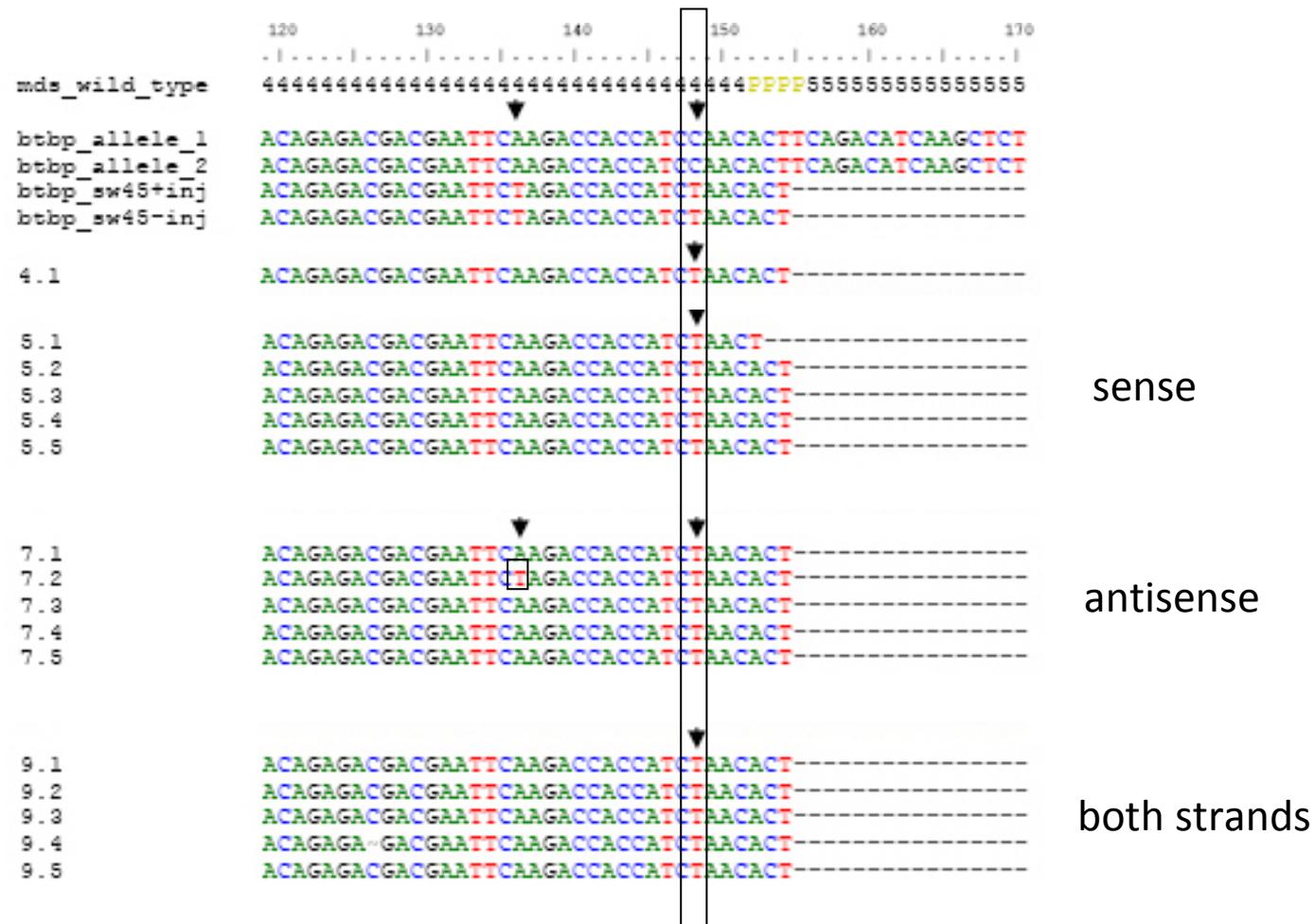
# Artificial RNA template microinjection re-programs the genome rearrangement pathway



# Switched chromosomes are transmitted to next generation



# Transfer of artificial nucleotide substitutions from RNA templates to nanochromosomes



# RNA-directed DNA proofreading

# Point substitutions are inherited by F2 and F3 generations

MDS 4 (MAC = Soma)

310 allele\_1 ...ATTCAAG...CATCCAACppp...  
310 allele\_2 ...ATTCAAG...CATCCAACppp...  
510 allele\_1 ...ATTCAAG...CATCCAACppp...  
510 allele\_2 ...ATTCAAG...CATCCAACppp...  
sw45 temp ...ATTCTAG...CATCTAACppp...

F1

...ATTCAAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...

310 allele\_1 ...ATTCAAG...CATCCAAC...  
310 allele\_2 ...ATTCAAG...CATCCAAC...  
510 allele\_1 ...ATTCAAG...CATCCAAC...  
510 allele\_2 ...ATTCAAG...CATCCAAC...  
sw45 temp ...ATTCTAG...CATCTAAC...

F1

...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...

F2

...ATTCAAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...

F2

...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...

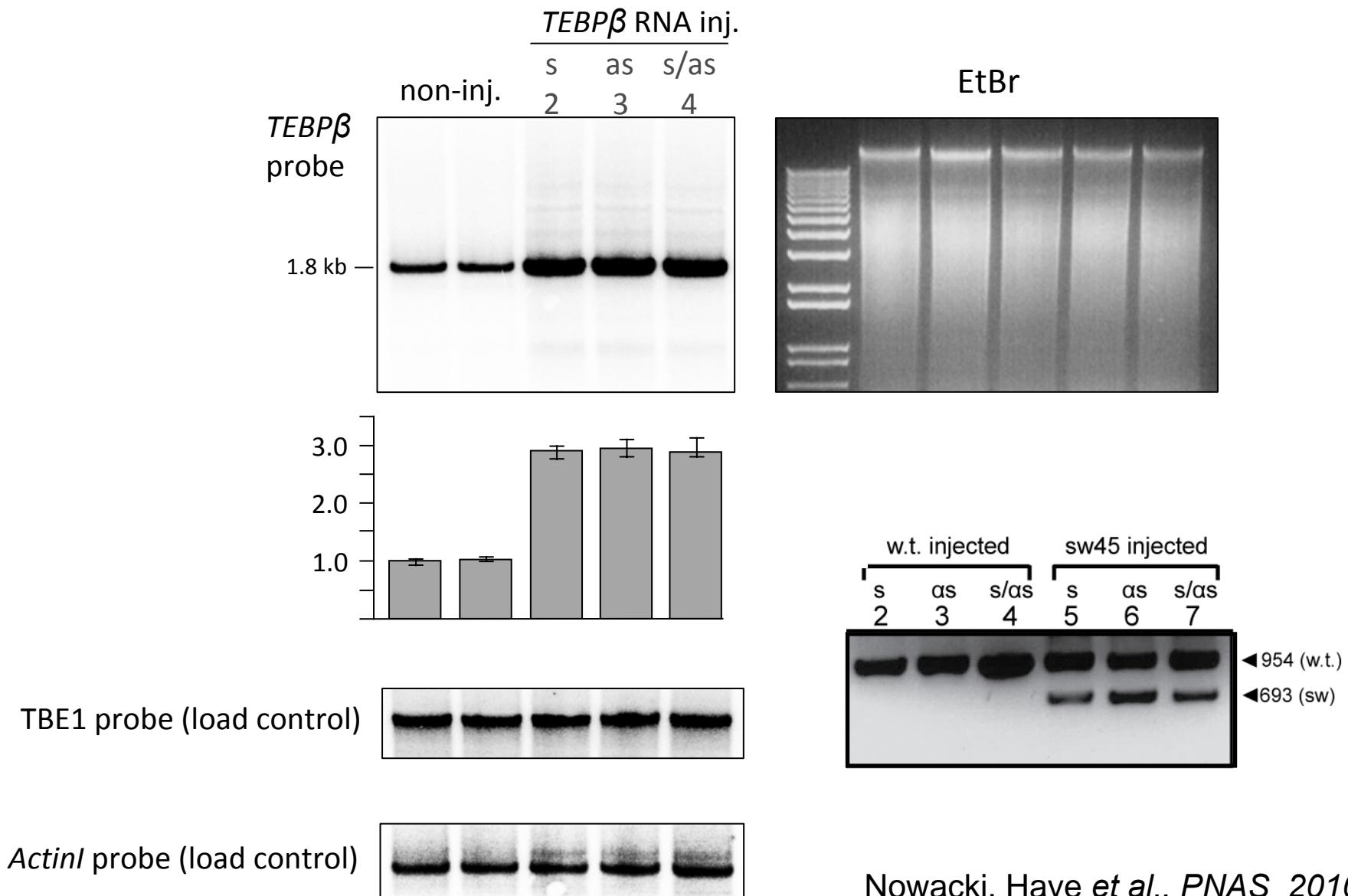
F3

...ATTCAAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...  
...ATTCTAG...CATCTAACppp...  
...ATTCAAG...CATCTAACppp...

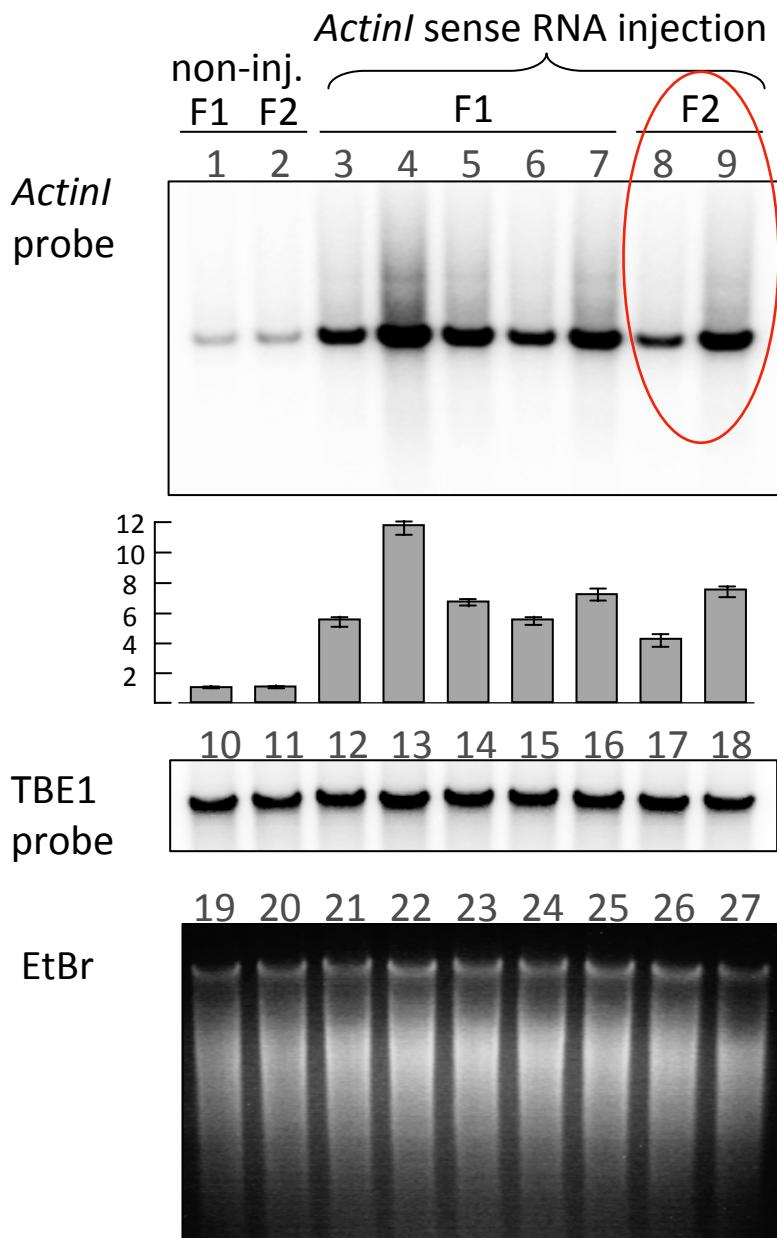
F3

...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...  
...ATTCAAG...CATCCAAC...

# Epigenetic changes in DNA copy number

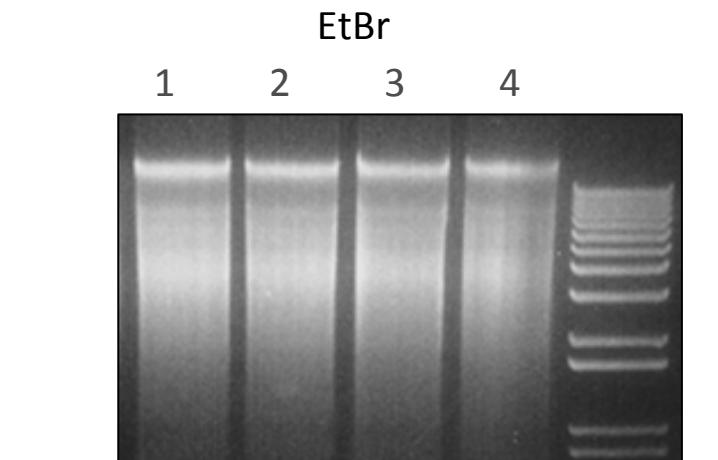
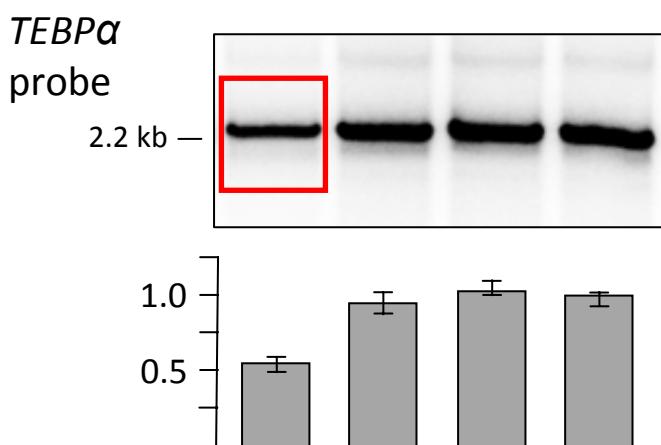
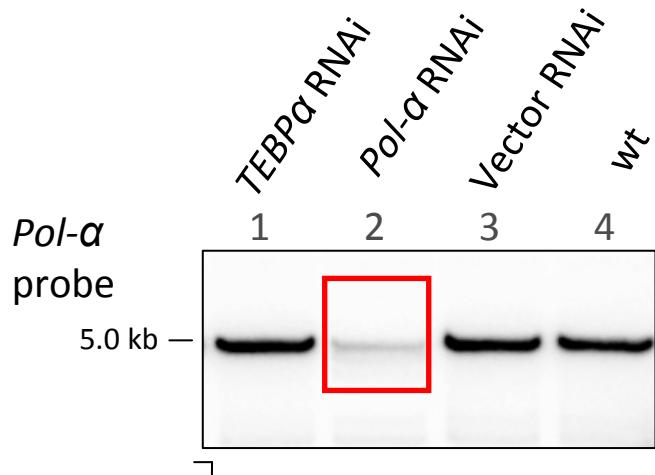


# Epigenetic inheritance of DNA copy number changes

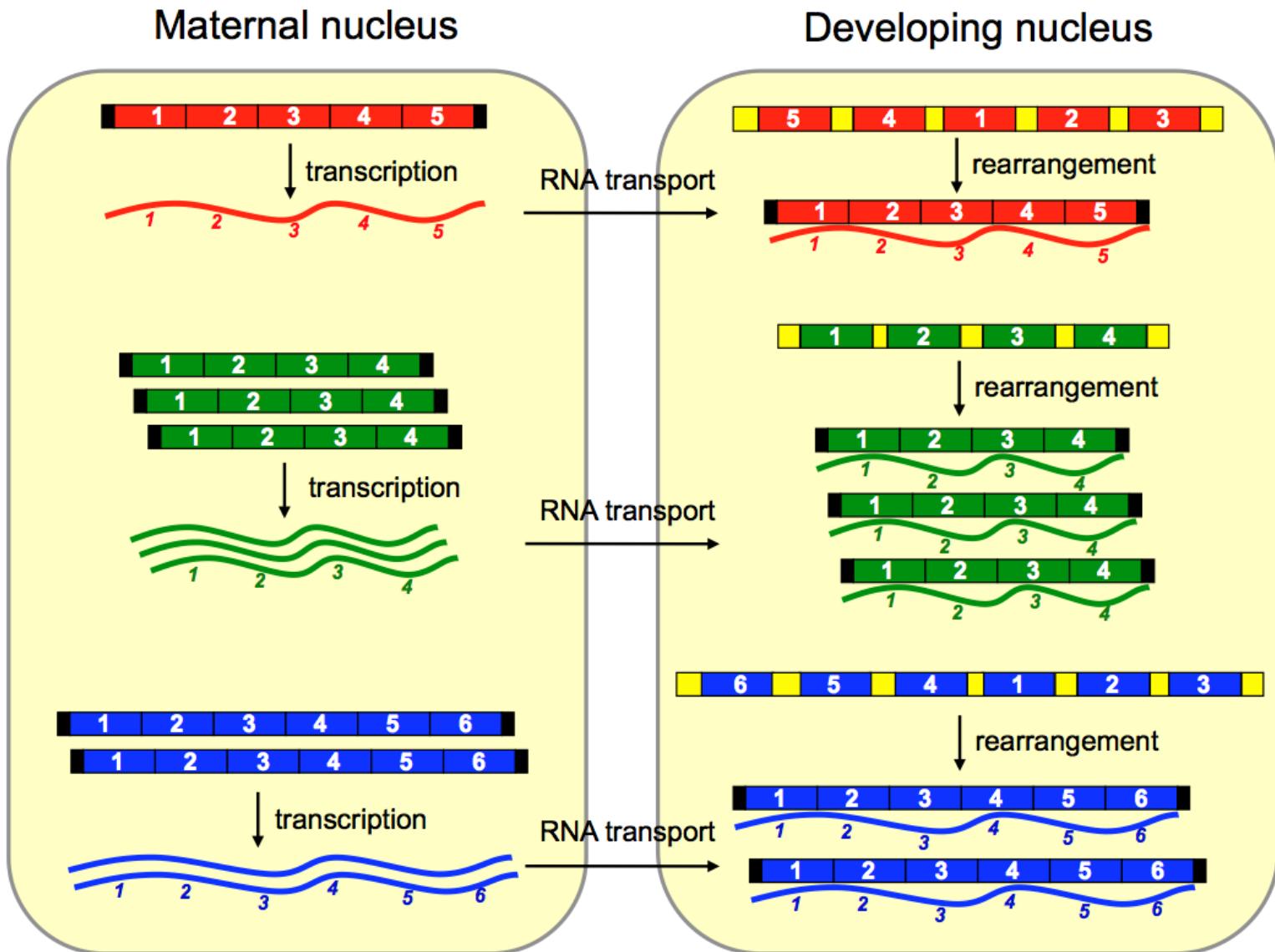


Nowacki, Haye et al.,  
PNAS 2010

# Reduced RNA template levels reduce DNA copy number



# Templates as epigenetic regulators of genome architecture



# Open Questions

- What polymerase makes templates?
  - How do RNA template-introduced mutations affect *Oxytricha* evolution?
- What machinery mediates the rearrangement process?
  - A role for transposases only found in the MIC genome (Nowacki *et al.*, 2009)
- How are deleted and retained regions specified?
  - A role for small RNAs (Fang *et al.*, in revision)

# Acknowledgements

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Xing Wang

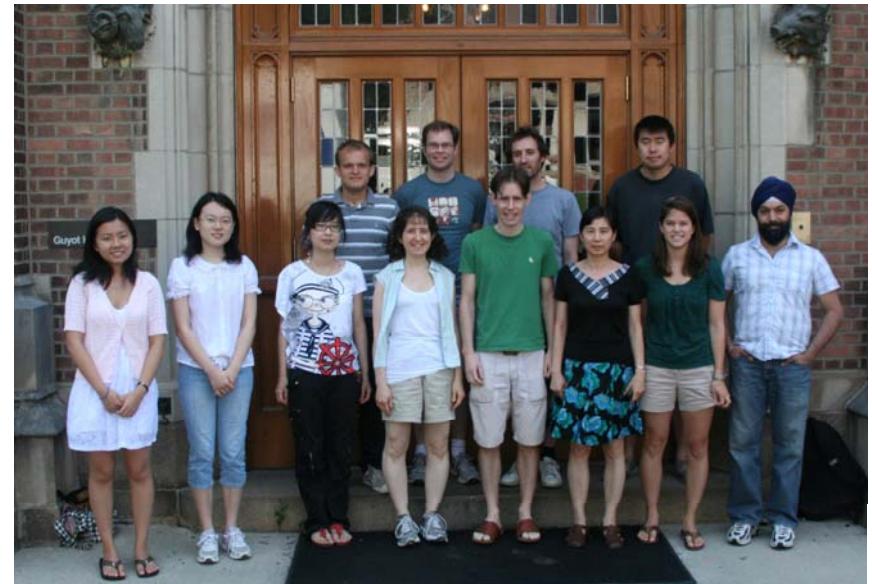
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**Benaryoya Research Institute**

Chris Amemiya

Andrew Stuart